



## 7. Southeast Atlantic and Gulf of Mexico Region Acidification Research

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### *Abstract*

The Southeast Atlantic and Gulf of Mexico Region encompasses continental shelf waters extending from the North Carolina to Florida coasts on the Atlantic seaboard and the marginal sea bounded by the U.S. Gulf Coast. While these two regions experience different stress factors with regards to ocean acidification (OA), they share similar needs with regards to local community engagement (or lack thereof), active research, and data availability. The regional influence of the Northward flowing Gulf Stream and southward flowing Labrador Sea currents in the Southeast Atlantic dominates the biogeochemical signatures of coastal waters in this region while the Gulf of Mexico is strongly influenced by the loop current and riverine inputs, which contribute to eutrophication and hypoxia. Impacts to coral reefs and the recreational and indus-

trial fishing industries, and potential prevalence and frequency of harmful algal blooms are some of the issues this region faces that are potentially affected by increasing ocean acidity. NOAA's Southeast Atlantic and Gulf of Mexico research goals are to:

- Expand OA monitoring using both traditional and new autonomous technologies to observe critical regions, including the ocean sub-surface and bottom water layer, to better characterize regional processes and improve fundamental understanding;
- Characterize ecosystem impacts and adaptive potential of species, with an aim to identify indicator species that can be used for early detection of unfavorable ecosystem conditions; and
- Use new knowledge to develop socioeconomic impact assessments of OA on recreation, tourism and aquaculture industries.

### *Acidification in the Atlantic and Gulf of Mexico Region*

The Gulf of Mexico (GoM) includes coastal areas of Florida (FL), Alabama (AL), Mississippi (MS), Louisiana (LA) and Texas (TX) and is a semi-enclosed marginal sea with coastal and open ocean waters. The eastern side of GoM includes the West Florida shelf, measuring up to 250 km while the Northern

and Western GoM shelves are much narrower. The Southeast Atlantic region (SE) is composed of the coastal areas of North and South Carolina, Georgia, and the East coast of Florida and is characterized by a shelf width on the order of tens of kilometers. It is bound by the Gulf Stream, which flows north-eastward along the shelf edge before detaching at Cape Hatteras, NC. The Gulf Stream is influenced by contributions from the SE and GoM region, although the influence of these regional inputs to OA variability has yet to be directly researched. Research described in this chapter excludes the Gulf of Mexico coral reefs in the keys and in the Flower Garden National Marine Sanctuary, which are instead addressed collectively with the Caribbean corals in *Chapter 8: Florida Keys and the Caribbean Region*.

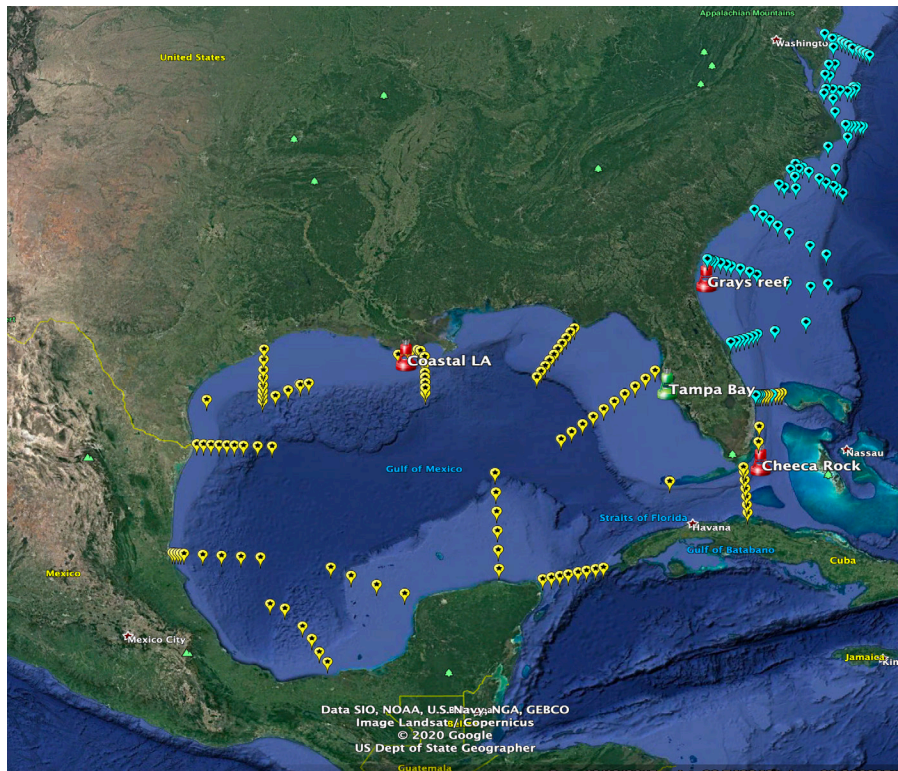
Slope water composition in the Southeast Atlantic region is a varying mix of predominantly Gulf Stream and Labrador Sea water, as well as inputs from coastal marshes. There is significant interannual variability in the region, primarily driven by the influence of different water masses, which affects OA conditions (Wang et al., 2013; Wanninkhof et al., 2015). Seasonal phytoplankton blooms do not occur regularly, and biologically driven carbon dioxide (CO<sub>2</sub>) uptake is less pronounced than in Atlantic coastal areas farther north (see *Chapter 9: Mid-Atlantic Bight Region* and *Chapter 10: New England Region*). The acidification rate in the South Atlantic Bight is higher than in the open ocean due to the combined effects of increased temperature in the middle and outer shelves, and lateral land-ocean interactions in the inner shelf (Reimer et al., 2017). Recent work using pH data collected for over a decade in two estuaries in North Carolina showed variations in pH linked to increasing river discharge and highlighted the importance of eutrophication (Van Dam & Wang, 2019). Dredging, water management, and associated activities in inlets and port areas (e.g., in Port Everglades) can cause underappreciated impacts on OA in South Florida. These activities can have coastal co-stressor effects (e.g., due to input from eutrophied, organic-rich freshwater canals and rivers) that can lead to enhanced acidification and impact local reefs and other organisms of economic interest (Enochs et al., 2019). This same area has had persistent harmful algal blooms (HABs) in

the past decades (Kramer et al., 2018), but studies relating them to OA have been inconclusive.

In the GoM, ocean water enters through the Yucatan channel and exits through the Florida Straits. Main features affecting water circulation in the GoM include the meandering Loop Current, which often sheds anticyclonic eddies that drift westward and can impact the shelf, and riverine input, particularly from the Mississippi-Atchafalaya river system, which provides large volumes of fresh water, nutrients and sediments. This riverine input can lead to eutrophication, hypoxia, and enhanced acidification (Cai et al., 2011). On the West Florida shelf riverine and groundwater input with high phosphate from natural deposits can have a unique signature of biologically mediated OA. Most of the coastal acidification studies in the GoM have focused in the northern and eastern coasts (e.g., Cai et al., 2011; Huang et al., 2013, 2015; Lohrenz et al., 2010; Feely et al., 2018; Hu et al., 2018; Robbins et al., 2018). Less data are available in the Western GoM shelf, with the exception of some estuarine work (McCutcheon et al., 2019) and very little data beyond surface measurements in deep waters of the GoM. The GoM also presents considerable regional variability. The West Florida Shelf exhibits supersaturated aragonite levels that vary between 2 and 5, both at surface and subsurface levels (Robbins et al., 2018) whereas the Northern GoM presents a steeper drop in saturation levels that is enhanced due to hypoxia (Cai et al., 2011; Feely et al., 2018). OA conditions in the GoM can vary significantly on an annual scale because of interannual variability in wind, temperature, precipitation, and water mass distributions (Muller-Karger et al., 2015; Wanninkhof et al., 2015).

Overall, the various observation- and model-derived estimates for the region agree in terms of their broad patterns, but there remain discrepancies between different estimates, which indicate that continued physical, chemical, and additional biological observational data in conjunction with modelling efforts are necessary in the region (e.g., Xue et al., 2016; Laurent et al., 2017; Lohrenz et al., 2018; Chen et al., 2019).





**Figure 7.1.** Location of NOAA-funded buoys (in red) in Gray’s Reef, Cheeca Rocks and Coastal Louisiana; non-NOAA funded buoy (in green) in Tampa Bay; and transects occupied as part of East Coast Ocean Acidification (ECO) and Gulf of Mexico Ecosystems and Carbon (GOMECC) research cruises (blue and yellow pins, respectively).

### *Environmental Change in the Southeast Atlantic and Gulf of Mexico Region*

Most OA observations in the Southeast and GoM coastal regions have focused on the chemical characterization of the area, particularly through NOAA and United States Geological Survey (USGS) efforts. NOAA-supported OA monitoring in the region currently includes: a) synoptic research cruises b) underway  $p\text{CO}_2$  systems installed on ships of opportunity, and c) coastal buoys (**Figure 7.1**). NOAA-led OA synoptic cruises have been carried out in the summertime every four years since 2007 in the East coast (East Coast OA (ECO) cruises) and the Gulf of Mexico (Gulf of Mexico Ecosystems and Carbon Cycle Cruise (GOMECC)) and they collect mostly physical and chemical data along a series of coastal transects (Wang et al., 2013; Wanninkhof et al., 2015). However, the GOMECC cruise (GOMECC-3), conducted in July-August of 2017, incorporated significant biological sampling in conjunction with OA water sampling, strengthening the link between OA forcing and impacts. NOAA-funded cruises in South Florida as part of the South Florida Ecosys-

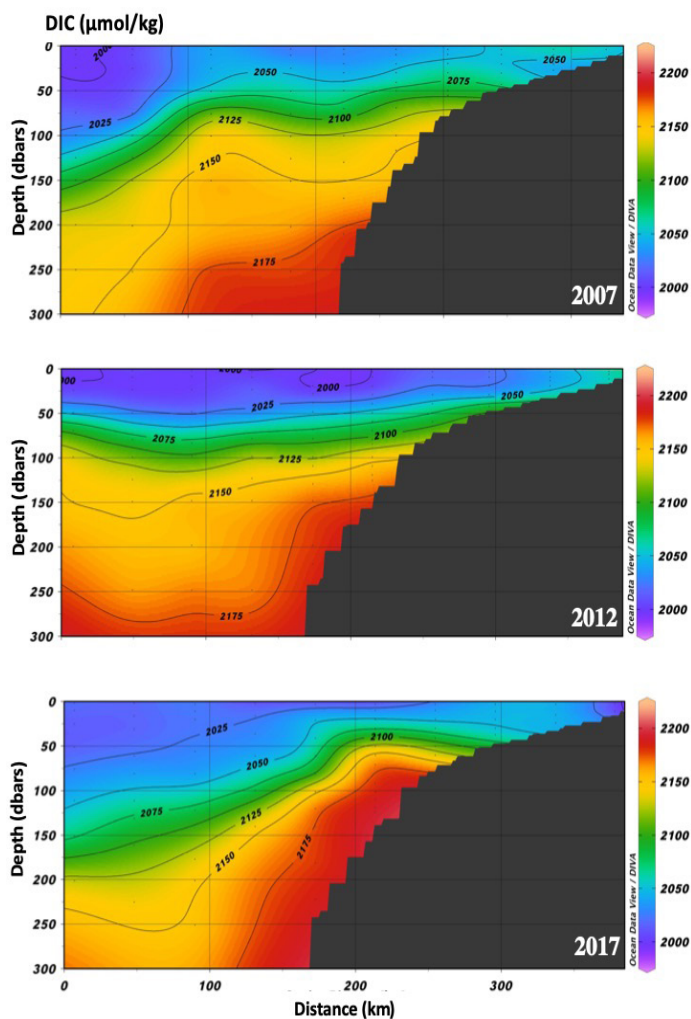
tem Restoration Program also collect OA samples along the FL keys and offshore of the Everglades several times per year. The USGS has participated in several cruises to study the effects of OA on marine organisms and habitats in the West Florida Shelf and northern Gulf of Mexico regions (Robbins et al., 2018). NOAA funds two underway  $p\text{CO}_2$  systems installed on NOAA fisheries ships RV *Gordon Guntter* and RV *Henry H. Bigelow* involved in regular fisheries surveys. In addition to these, NOAA supports underway  $p\text{CO}_2$  systems through its Ship of Opportunity Program (SOOP). These systems are installed on a variety of ships including scientific and commercial vessels that have also collected data in the Southeast and Gulf of Mexico regions. As a result, over 90% of available surface  $p\text{CO}_2$  data in the Gulf of Mexico has been collected through NOAA-funded efforts since 2008. Surface water samples for carbonate chemistry analysis are also collected on these ships on an opportunistic basis. Additionally, ships of opportunity provide platforms for maintenance of three NOAA OA monitoring buoys in the region (Sutton et al., 2019), and collect data for biogeochemical modeling efforts. The buoys are locat-

ed off the Georgia coast in Gray's Reef ( $p\text{CO}_2$  record started in 2006, pH sensor added in 2011), in the Florida Keys at Cheeca Rocks ( $p\text{CO}_2$  and pH record started in 2011) and in coastal Louisiana ( $p\text{CO}_2$  and pH record available since 2011). A fourth OA buoy is located in Tampa Bay, FL, and is maintained by Dr. Kim Yates, from the USGS. **Figure 7.1** shows the location of the buoys as well as the transects occupied in the ECOA and GOMECC cruises. Although none are in use in the region, a variety of autonomous platforms equipped with a variety of OA sensors are being explored in other regions. Examples of such platforms include wave gliders for surface measurements, saildrones and BGC-Argo profiling floats, which could provide an excellent means to increase data availability in areas such as the deep GoM.

Observing capabilities over the last decade have greatly improved the spatial coverage of carbonate chemistry measurements made in the region and contributed toward the foundational understanding of the large-scale regional trends from different source waters in the region, and the impacts of riverine inputs and eutrophication on OA conditions in the region. Quantifying and understanding the high natural variability, and teasing apart anthropogenic impacts from natural variability, including from water mass composition, biological activity, and river discharge remain research foci of OA drivers and co-stressors in the region.

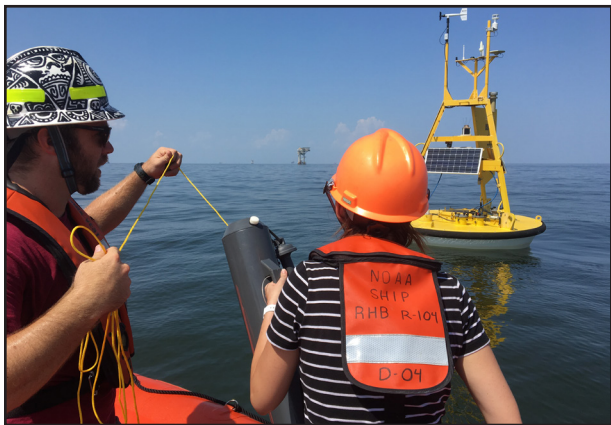
The ECOA and GOMECC cruises as well as several other OA cruises in the Northern GoM collect high-resolution water column data, and take place during the summer season in order to provide an interannual comparison without confounding factors from seasonal variability. As a result there are insufficient observations from other seasons to adequately characterize seasonal variability in subsurface waters and the open ocean end-member (**Figure 7.2**). Moreover, there are presently no OA buoys in the Western GoM, representing a major regional sampling gap.

Nearshore estuarine and coastal regions in the GoM and Southeastern Atlantic have also been relatively undersampled. This includes mangroves, marshes, and some estuaries that provide a wealth



**Figure 7.2.** Concentration of dissolved inorganic carbon (DIC) off the West Florida shelf measured during the Gulf of Mexico Ecosystems and Carbon Cruise (GOMECC). Increased concentrations are observed in the subsurface over time. Surface measurements cannot capture changes in subsurface waters.

of ecosystem services in this region, but are also data poor environments for OA. The majority of recreational fishing and tourism occurs in these data poor nearshore waters; thus, increased sampling is commencing for nearshore waters. The GoM and Southeast Atlantic region have a combined 22 National Parks that have observational infrastructure, support, and outreach opportunities. Of these, 7 have started collecting OA samples in coordination with the synoptic GOMECC and ECOA cruises, once every 4 years. There are two National Marine Sanctuaries (NMS) in the GoM, the Florida Keys NMS and the Flower Garden Banks NMS that are coral reef areas heavily instrumented for OA research and monitoring (see Chapter 8: Florida Keys and Caribbean Region). However, there are no similar monitoring efforts in place for cold-water corals in the Gulf of Mexico, which live at depths between 300 and 600 m, and which have been identified as being particularly vulnerable communities to OA because they are already naturally exposed to lower pH levels (Lunden et al., 2013; Georgian et al., 2016). In the Southeast Atlantic region Gray's Reef NMS off the Georgia coast has a buoy with OA-monitoring systems.



NOAA researchers collect a water sample near an acidification buoy during a Gulf of Mexico/East Coast Carbon research cruise. Credit: Leticia Barbero/NOAA

The in-situ observing data in the region are used in existing modeling efforts including the OA Product Suite developed for the Greater Caribbean region and now extended to include the East Coast. The OA Product Suite utilizes satellite data and a data-assimilative hybrid model to produce monthly maps of surface water carbonate system components. Modeling results include studies in the Gulf of Mex-

ico by Xue et al. (2016), who showed the GoM to be an annual sink for CO<sub>2</sub> with a flux of  $1.11 \pm 0.84 \cdot 10^{12}$  mol C yr<sup>-1</sup>, and by Laurent et al. (2017), who showed the recurring development of an extended area of acidified bottom waters in summer on the Northern GoM shelf that coincides with hypoxic waters. Global data synthesis and process-based global models have also provided estimates for coastal U.S. regions including the GoM and Southeast (see Table 1 in Fennel et al., 2019). More recently, modeling efforts are being supported to address seasonal patterns of the carbon system. Modeling results agree in general terms with observation estimates, and offer the opportunity to upscale the observations in time and space. However, the models still show significant discrepancies in some regions, specifically in the period from 2010 to 2017 depending on the model. Sustaining current modeling efforts and increasing the modeling portfolio to include nowcasts and forecasts, as well as integrating ecosystem parameters, extending to the East coast, and including robust validation is a priority to predict the expected changes in the region as a result of OA.

### ***Research Objective 7.1: Improve characterization of OA parameters in important economic, cultural, and recreational regions***

The GoM and Southeast Atlantic are particularly data poor in the nearshore region where most commercial and recreational fishing and tourist industry activities take place. These regions are also home to mangroves, marshes, estuaries, natural, and restored oyster reef sites, which are essential habitats within the region's marine ecosystems, play an important role in local carbon balances, and provide a wealth of ecosystem and commercial services.

**Action 7.1.1:** Develop and establish protocols that are complementary to ongoing synoptic cruises to extend regular observations of pertinent OA parameters into the nearshore environment. Focus sampling in Essential Fish Habitats for species predicted to be significantly impacted by OA, and in select National Parks and National Marine Sanctuary sites (specifically in the northern GoM where hypoxia and OA act as co-stressors).



**Action 7.1.2:** Explore options, including private and/or industry partnerships, to add an OA buoy or alternative observing platform in the Western GoM to extend coverage of coastal sites that are presently composed of buoys in the Florida Keys, West Florida Shelf (a non-NOAA asset), and the Mississippi-Atchafalaya area.

**Action 7.1.3:** Explore options to add a monitoring site in the SE region within the estuarine environment near to Grays Reef to establish a nearshore-offshore contrasting monitoring site.

**Action 7.1.4:** Establish OA and water quality monitoring stations at inlets and near commercially and recreationally important estuaries (e.g., oyster bed leases, public clam beds, shellfish hatcheries) to monitor coastal acidification and eutrophication co-stressors in areas where fresh water systems are highly impacted by human activities and strongly influence coastal oceans.

### ***Research Objective 7.2: Improve the characterization of the Open Ocean***

Currently, open ocean monitoring is limited to the OA synoptic cruises that take place once every four years. Exploring autonomous technologies may provide a platform to vastly increase open ocean observing in the deep and shelf waters. Better understanding of the open ocean within the region can also contribute to the understanding of coastal acidification processes, as coastal acidification can be studied by considering a conservative mixing line with a two end-member system of open-ocean and freshwater inputs.

**Action 7.2.1:** Evaluate capabilities of autonomous sensor(s) for surface to deep water observing (3000 m) and observing in the vicinity of cold-water coral communities.

**Action 7.2.2:** Establish plan to deploy BGC-Argo floats in the GoM region, following the rationale in *Chapter 2: Open Ocean Region (Objective 2.3)*. Leverage GOMECC and other cruises to perform *in-situ* calibrations and improve quality control procedures for the data, while greatly increasing data availability for the open ocean end-member in the GoM.

### ***Research Objective 7.3: Improve fundamental understanding of regional processes and seasonal trends***

Coastal areas in the region show different seasonal surface and sub-surface patterns, but there is little data available to validate model estimates of spatial patterns and seasonal trends.

**Action 7.3.1:** Leverage existing cruises (e.g., Southeast Area Monitoring and Assessment Program, ecosystem monitoring and restoration, oceanographic) to increase sample collection between synoptic surveys, particularly in wintertime when observations have historically been particularly limited.

**Action 7.3.2:** Evaluate methods to measure how upwelling of deep GoM waters onto the shelf affect OA in shelf ecosystems that are also affected by riverine acidification impacts.

**Action 7.3.3:** Expand the number of observations by increasing frequency of synoptic cruises to sample during other seasons (initially winter) to improve intra-annual sampling and add subsurface sensors to existing buoys, moorings, and autonomous platforms.

### ***Research Objective 7.4: Improve scaling and predictive capabilities***

Models are a critical tool for extrapolating current observations to larger regions and to improve our mechanistic understanding of linkages between the physics, chemistry, and biology of OA. Models also provide critical information that can be translated and used to inform decisions and management practices. Continued development and use of existing regional models and the creation of new models that enhance geographic coverage within the region are needed.

**Action 7.4.1:** Develop, apply, and improve existing models and validate models with direct observations to assess and improve model skill to best project OA within the region.

**Action 7.4.2:** Incorporate OA and associated biogeochemistry into ecosystem models to help predict OA impacts on valuable components of the marine ecosystem.

**Action 7.4.3:** Increase utilization of satellite data, tools, and products in support of status estimates and now-casts.

**Action 7.4.4:** Coordinate research with university researchers to build consensus regarding regional OA projections.

### **Biological Sensitivity**

Studies of OA impacts on organisms in the region, with the exception of some coral reef systems in the Florida Keys and Flower Garden NMS (Chapter 8), have been sparse. Changes in chemical factors directly impact the physiology of filter feeders, benthic foragers and fish (e.g., oysters, blue crab or menhaden) and also alter food web structure and the quality and quantity of food for many of the intermediate consumers and commercially important species (Hansen et al., 2019; Caron & Hutchins, 2012). Initial characterizations of food web structure - from phyto- to zooplankton - were performed during the 2017 and 2018 synoptic OA GOMECC-3 and ECOA-2 cruises across the GoM and Southeast Atlantic region. GOMECC-3 also included targeted sampling for pteropods and larval ichthyoplankton, along with rate measurements of carbon flow from primary producers to crustacean prey. Incorporation of rate-based measurements, specifically, allow for more investigation of the role of these biological communities and food webs as sinks of or links to carbon cycling in the region (Sherr & Sherr, 2002; Steinberg & Landry, 2017). Altogether, this information provides insights into how food webs and carbon transfer are altered in regions affected by eutrophication-driven acidification and hypoxia. Maps of ichthyoplankton distribution are reflective of spawning regions for fish, and can be used for studying effects of OA on marine fish populations such as stock displacement to avoid acidified waters. These data build directly on programs such as Southeast Area Monitoring and Assessment Program (SEAMAP) and would provide more power to efforts to depict changes in species ranges and patterns of distribution during the larval life stages and beyond.

Given the lack of a systematic study of OA impacts on plankton and commercially important species in this region, impacts of OA on fisheries and

aquaculture industries in the region are also poorly understood. With high saturation state variability predicted for subtropical regions relative to higher latitudes, understanding the impacts of saturation state on such species will be critically important for the region. High levels of OA variability could lead to greater impacts on organisms that have a smaller tolerance to OA. Identifying such highly OA-sensitive indicator species could actually be beneficial to tracking small and/or early shifts in the marine carbonate system. The Southeast Fisheries Science Center (SEFSC) has conducted a climate vulnerability assessment for marine fishery species in the GoM and initiated one in the SE (Lovett et al., 2016), which can guide the proposed *in-situ* observing activities (*Research Objective 7.3*) and indicator species research (*Research Objective 7.6*) proposed for the region.

Oil drilling, dredging and restoration efforts both in the GoM and SE region can interact with OA and potentially have compounding impacts on organisms and ecosystems. While funding is available for research on OA, oil spills, or hypoxia, there are no coordinated calls to promote and encourage interdisciplinary research. HAB events also pose a recurring problem in the GoM and Southeast region. Florida and other Gulf Coast states experience fish kills and neurotoxic shellfish poisoning from *Karenia brevis* and other HAB species (Weisberg et al., 2019). Harmful cyanobacteria and their toxins (primarily microcystin) have also been detected in low salinity estuaries in Louisiana and Florida (Bargu et al., 2011; Riekenberg et al., 2015) and shown to accumulate in commercially-important consumer species (i.e., blue crab) that are also impacted by OA (Garcia et al., 2010). The economic impact of HABs resulting in public health issues, commercial fishery closures, and recreational tourism reduction has been reported as upwards of ~\$50 million/year (Anderson et al., 2000). Laboratory studies of *Karenia brevis*, the major HAB species in the Gulf of Mexico, in connection with OA have shown conflicting results, with some concluding that at higher  $p\text{CO}_2$  concentrations *K. brevis* growth rates are significantly increased, although toxin production itself appeared to not be linked (Errera et al., 2014), while others did not observe a significant response in growth, or cellular composition of carbon and ni-

trogen (Bercel & Kranz, 2019). Although ongoing efforts to study the relationship between OA and HAB occurrence are taking place in other regions, no similar effort is currently taking place in the GoM.



Small boat operations as part of the Gulf of Mexico Ecosystems and Carbon Cruise aboard the NOAA ship Ronald H. Brown. Credit Marisa Gedney/NOAA

### ***Research Objective 7.5: Increase understanding of the impacts of OA on ecosystem productivity and food webs***

Because plankton communities are the base of marine food webs, shift in response to OA impact energy flow and ecosystem function (Roman et al., 2012). As the quantity and quality of plankton prey are altered, managed and commercially important species are affected.

**Action 7.5.1:** Characterize plankton communities (from phytoplankton to larval fish) along spatial gradients of eutrophication-driven acidification and hypoxia through regular sampling on GOMECC, ECOA, and other cruises to allow for attribution to OA and/or eutrophication stressors versus seasonal or other episodic drivers (e.g., tropical storms, flood, drought).

**Action 7.5.2:** Quantify changes in carbon flow to higher trophic levels (e.g., crustaceans and fish) via modeling studies and shipboard observations during GOMECC and ECOA cruises. Conduct shipboard experiments to determine biological community composition during antecedent conditions (not just conditions at the time of sampling) and to understand how rates (e.g., primary productivity, zooplankton grazing) change in response to OA, eu-

trophication, HABs, and hypoxia, which are critical to parameterize ecosystem models.

**Action 7.5.3:** Synthesize existing information from previous cruises and ongoing research and monitoring in the region. Coordinate collection of biological data (e.g., plankton tows, 'omics-approaches and rate measurements) for future GOMECC, ECOA, and other cruises. Identify regions where shifts in carbon chemistry are associated with changes in plankton community structure and function.

### ***Research Objective 7.6: Identify indicator species for OA in the region***

An indicator species that is sensitive to changes in pH specific for the GoM and for the Southeast Atlantic region can be used for early detection of OA impacts to the system and to investigate ecosystem impacts that may result from food web changes.

**Action 7.6.1:** Incorporate plankton and neuston net tows, and 'omics sampling as part of the standard suite of parameters included in GOMECC/EOA cruises.

**Action 7.6.2:** Incorporate carbon chemistry sampling as part of the standard suite of parameters included in already ongoing SEFSC ecosystem monitoring efforts such as SEAMAP cruises and add DIC/TA/pH water sampling to the suite of samples already being collected.

**Action 7.6.3:** Conduct laboratory studies to examine OA impacts in combination with other co-stressors, such as temperature and nutrients, on potential indicator species identified via field observations.

### ***Research Objective 7.7: Characterize sensitivity and adaptive potential of critical resource species to OA and other stressors and improve the understanding of OA impacts to HAB event frequency and duration***

Most species of economic interest in the region (e.g., bluefin tuna, shrimp, blue crab) lack specific studies about potential OA impacts. The SEFSC vulnerability analysis mentioned above and 'omics tools can be used as a screening tool to identify species of



economic importance that are likely sensitive to OA. In addition to this, a growing body of research is addressing whether OA may have species-specific impacts on the frequency, duration and degree of HAB blooms or their toxicity in other regions of the U.S. Despite the prevalence of HABs, research focused on GoM and Southeast Atlantic environments and species with regards to OA is scarce.

**Action 7.7.1:** Target species of interest to conduct experimental studies to establish responses to OA and inform species vulnerability assessments.

**Action 7.7.2:** Develop assessments using a multi-stressor framework to include combinations of effects such as eutrophication, river runoff, hypoxia, or increased HABs.

**Action 7.7.3:** Incorporate these results into ecosystem models to drive hypotheses about how changes in indicators species and plankton dynamics will affect commercial and recreational fishery species.

**Action 7.7.4:** Build monitoring capacity for regionally significant HAB species to be measured during synoptic OA cruises and implement OA sampling in other opportunistic or ongoing cruises organized in relation to HABs already occurring along the Florida coast.

**Action 7.7.5:** Support isolation and cultivation-based laboratory experimentation of local HAB species to examine species-specific and community responses to carbonate chemistry conditions.

**Action 7.7.6:** Quantify socioeconomic impacts from predicted changes in HABs and their toxicity due to OA.

### **Human dimensions**

While there are several studies that deal with socioeconomic impacts as a result of acidification in coral reef regions, there are currently no non-coral studies in the GoM and the Southeast region that quantify potential socioeconomic impacts from OA on fisheries of interest in the region. Ekstrom et al. (2015) identified the coastal communities of TX, LA, MS and Northern FL as being highly socially vulner-

able to OA impacts, despite a lower marine ecosystem exposure than other U.S. coastal regions. However, no specific socioeconomic studies have been released yet. Local communities in the SE such as those of Gullah/Geechee descent consume marine species vulnerable to OA (e.g., oysters, blue crab) for sustenance and as part of cultural traditions. This type of activity could be included as part of the evaluation and research of socioeconomic impacts as an opportunity to build relationships and partnerships with vulnerable communities. Stakeholder engagement efforts should be cognizant that the region experiences threats from multiple drivers, some of which have a more direct impact than OA. Therefore efforts to engage stakeholders and identify needs should take a multi-stressor approach. By improving the understanding of stakeholder needs strategic investments can be made in capacity building and targeted research that is responsive to the needs within the region.



A commercial shrimp fishing vessel off of the U.S. southeast coast. Credit: NOAA

### **Research Objective 7.8: Improve assessment of socioeconomic impacts of OA on local tourism, recreational fishing, commercial fishing, and aquaculture (shellfish, fisheries) industries**

To date, no socioeconomic studies have been conducted to quantify the impact OA might have on commercially relevant fisheries, aquaculture, tourism, or recreational fishing in the region.

**Action 7.8.1:** Evaluate the socioeconomic impacts from from key species being impacted by OA either

directly or in through food web interactions (*Research Objective 7.7*).

**Action 7.8.2:** Conduct socioeconomic research to quantify the impacts of OA for specific fisheries, including direct (fishermen/aquaculture) and indirect (related service industries) impacts.

**Action 7.8.3:** Based on outcomes from above, involve local stakeholders and raise awareness about OA to increase community resilience and proactively develop OA mitigation plans for affected ecosystems, industries, and economies.

# References

[INTRO] = Introduction;

[NAT] = National Ocean & Great Lakes;

[OO] = Open Ocean Region;

[AK] = Alaska Region;

[ARC] = Arctic Region;

[WC] = West Coast Region;

[PAC] = U.S. Pacific Islands Region;

[SAG] = Southeast Atlantic & Gulf of Mexico Region;

[FLC] = Florida Keys and Caribbean Region;

[MAB] = Mid-Atlantic Bight Region;

[NE] = New England Region;

[GL] = Great Lakes Region

Aguilera, S. E., Cole, J., Finkbeiner, E. M., Le Cornu, E., Ban, N. C., Carr, M. H., et al. (2015). Managing small-scale commercial fisheries for adaptive capacity: insights from dynamic social-ecological drivers of change in Monterey Bay. *PLoS ONE*, 10(3), e0118992. <https://doi.org/10.1371/journal.pone.0118992> [WC]

Ainsworth, C. H., Samhouri, J. F., Busch, D. S., Chueng, W. W. L., Dunne, J., & Okey, T. A. (2011). Potential impacts of climate change on northeast Pacific marine fisheries and food webs. *ICES Journal of Marine Science*, 68(6), 1217–1229. [WC]

Alin, S., Brainard, R., Price, N., Newton, J., Cohen, A., Peterson, W., et al. (2015). Characterizing the natural system: Toward sustained, integrated coastal ocean acidification observing networks to facilitate resource management and decision support. *Oceanography*, 28(2), 92–107. [WC]

Alvarez-Filip, L., Dulvy, N. K., Gill, J. A., Cote, I. M., & Watkinson, A. R. (2009). Flattening of Caribbean coral reefs: region-wide declines in architectural complexity. *Proceedings of the Royal Society B*, 276(1669), 3019–3025. <https://doi.org/10.1098/rspb.2009.0339> [FLC]

AMAP (2013). AMAP Assessment 2013: Arctic Ocean Acidification. Oslo, Norway: Arctic Monitoring and Assessment Programme (AMAP). 99 pp. [ARC]

AMAP (2018). AMAP Assessment 2018: Arctic Ocean Acidification. Tromsø, Norway: Arctic Monitoring and Assessment Programme (AMAP), 187 pp. [ARC]

American Sportfishing Association (2013). American Sportfishing in America: An Economic Force for Conservation, Alexandria, VA. Retrieved from [http://asafishing.org/uploads/2011\\_ASASportfishing\\_in\\_America\\_Report\\_January\\_2013.pdf](http://asafishing.org/uploads/2011_ASASportfishing_in_America_Report_January_2013.pdf) [GL]

Anderson, D. M., Hoagland, P., Kaoru, Y., & White, A. W. (2000). Estimated annual economic impacts from harmful algal blooms (HABs) in the United States. (No. WHOI-2000-11). Norman, OK: National Oceanic and Atmospheric Administration, National Severe Storms Laboratory. [SAG]

Anderson, L. G., Tanhua, T., Björk, G., Hjalmarsson, S., Jones, E. P., Jutterström, S., et al. (2010). Arctic ocean shelf-basin interaction: An active continental shelf CO<sub>2</sub> pump and its impact on the degree of calcium carbon solubility. *Deep Sea Research Part I*, 57(7), 869–879. <https://doi.org/10.1016/j.dsr.2010.03.012>. [ARC]

Azetsu-Scott, K., Clarke, A., Falkner, K., Hamilton, J., Jones, E. P., Lee, C., et al. (2010). Calcium carbonate saturation states in the waters of the Canadian Arctic Archipelago and the Labrador Sea. *Journal of Geophysical Research*, 115, C11021. <https://doi.org/10.1029/2009JC005917> [ARC]

Barange, M., Merino, G., Blanchard, J. L., Scholtens, J., Harle, J., Allison, E. A., et al. (2014). Impacts of climate change on marine ecosystem production in societies dependent on fisheries. *Nature Climate Change*, 4, 211–216. <https://doi.org/10.1038/NCLIMATE2119> [AK]

Barange, M., Bahri, T., Beveridge, M. C. M., Cochrane, K. L., Funge-Smith, S., & Poulain, F. (Eds.) (2018). Impacts of climate change on fisheries and aquaculture: Synthesis of current knowledge, adaptation and mitigation options. *FAO Fisheries and Aquaculture Technical Paper*,



627. Rome, Italy: Food and Agricultural Organization of the United Nations, 628 pp. [AK]
- Bargu, S., White, J. R., Li, C., Czubakowski, J., & Fulweiler, R. W. (2011). Effects of freshwater input on nutrient loading, phytoplankton biomass, and cyanotoxin production in an oligohaline estuarine lake. *Hydrobiologia*, 661(1), 377–389. <https://doi.org/10.1007/s10750-010-0545-8> [SAG]
- Barkley, H. C., Cohen, A. L., Golbuu, Y., Starczak, V. R., DeCarlo, T. M., & Shamberger, K. E. (2015). Changes in coral reef communities across a natural gradient in seawater pH. *Science Advances*, 1(5), e1500328. <https://doi.org/10.1126/sciadv.1500328> [INTRO]
- Barton, A., Hales, B., Waldbusser, G. G., Langdon, C., & Feely, R.A. (2012). The Pacific oyster, *Crassostrea gigas*, shows negative correlation to naturally elevated carbon dioxide levels: Implications for near-term ocean acidification effects. *Limnology and Oceanography*, 57, 698–710. <https://doi.org/10.4319/lo.2012.57.3.0698> [WC]
- Barton, A., Waldbusser, G. G., Feely, R. A., Weisberg, S. B., Newton, J. A., Hales, B., et al. (2015). Impacts of coastal acidification on the Pacific Northwest shellfish industry and adaptation strategies implemented in response. *Oceanography*, 28(2), 146–159. <https://doi.org/10.5670/oceanog.2015.38> [WC]
- Bates, N. R. (2015). Assessing ocean acidification variability in the Pacific-Arctic Region as part of the Russian-American Long-term Census of the Arctic (RUSALCA). *Oceanography*, 28(3), 36–45. <https://doi.org/10.5670/oceanog.2015.56> [ARC]
- Bates, N. R., & Mathis, J. T. (2009). The Arctic Ocean marine carbon cycle: Evaluation of air-sea CO<sub>2</sub> exchanges, ocean acidification impacts, and potential feedbacks. *Biogeosciences*, 6, 2433–2459. <https://doi.org/10.5194/bg-6-2433-2009> [ARC]
- Bates, N. R., Cai, W.-J., & Mathis, J. T. (2011). The ocean carbon cycle in the western Arctic Ocean: Distributions and air-sea fluxes of carbon dioxide. *Oceanography*, 34(3), 186–201. [ARC]
- Bates, N. R., Astor, Y. M., Church, M. J., Currie, K., Dore, J. E., González-Dávila, M., et al. (2014a). A time-series view of changing ocean chemistry due to ocean uptake of anthropogenic CO<sub>2</sub> and ocean acidification. *Oceanography*, 27, 126–141. <https://doi.org/10.5670/oceanog.2014.16> [OO][PAC]
- Bates, N. R., Garley, R., Frey, K. E., Shake, K. L., & Mathis, J. T. (2014b). Sea-ice melt CO<sub>2</sub>-carbonate chemistry in the western Arctic Ocean: meltwater contributions to air-sea CO<sub>2</sub> gas exchange, mixed-layer properties, and rates of net community production under sea ice. *Biogeosciences*, 11, 6769–6789. <https://doi.org/10.5194/bg-11-6769-2014> [ARC]
- Bauer, M., Hoagland, P., Leschine, T. M., Blount, B. G., Pomeroy, C. M., Lampl, L. L., et al. (2010). The importance of human dimensions research in managing harmful algal blooms. *Frontiers in Ecology and the Environment*, 8(2), 75–83. <https://doi.org/10.1890/070181> [GL]
- Baumann, H. (2019). Experimental assessments of marine species sensitivities to ocean acidification and co-stressors: How far have we come? *Canadian Journal of Zoology*, 97(5), 399–408. <https://doi.org/10.1139/cjz-2018-0198> [NE]
- Baumann, H., Talmage, S. C., & Gobler, C. J. (2012). Reduced early life growth and survival in a fish in direct response to increased carbon dioxide. *Nature Climate Change*, 2(1), 38–41. <https://doi.org/10.1038/nclimate1291> [NE]
- Beare, D., McQuatters-Gollop, A., van der Hammen, T., Machiels, M., Teoh, S. J., & Hall-Spencer, J. M. (2013). Long-term trends in calcifying plankton and pH in the North Sea. *PLoS ONE*, 8, e61175. <https://doi.org/10.1371/journal.pone.0061175> [INTRO]
- Bednaršek, N., & Ohman, M. (2015). Changes in pteropod distributions and shell dissolution across a frontal system in the California Current System. *Marine Ecology Progress Series*, 523, 93–103. <https://doi.org/10.3354/meps11199> [WC]
- Bednaršek, N., Feely, R. A., Reum, J. C. P., Peterson, B., Menkel, J., Alin, S. R., & Hales, B. (2014). *Limacina helicina* shell dissolution as an indicator of declining habitat suitability owing to ocean acidification in the California Current Ecosystem. *Proceedings of the Royal Society B: Biological Sciences*, 281(1785), 20140123. <https://doi.org/10.1098/rspb.2014.0123> [OO][WC]
- Bednaršek, N., Harvey, C. J., Kaplan, I. C., Feely, R. A., & Možina, J. (2016). Pteropods on the edge: Cumulative effects of ocean acidifica-

- tion, warming, and deoxygenation. *Progress in Oceanography*, 145, 1–24. <https://doi.org/10.1016/j.pocean.2016.04.002> [WC]
- Bednaršek, N., Feely, R. A., Tolimieri, N., Hermann, A. J., Siedlecki, S. A., Waldbusser, G. G., et al. (2017a). Exposure history determines pteropod vulnerability to ocean acidification along the US West Coast. *Scientific Reports*, 7(1), 4526. <https://doi.org/10.1038/s41598-017-03934-z> [OO][WC]
- Bednaršek, N., Klinger, T., Harvey, C. J., Weisberg, S., McCabe, R. M., Feely, R. A., et al. (2017b). New ocean, new needs: Application of pteropod shell dissolution as a biological indicator for marine resource management. *Ecological Indicators*, 76, 240–244. [INTRO][WC]
- Bednaršek, N., Feely, R. A., Beck, M. W., Glippa, O., Kanerva, M., & Engström-Öst, J. (2018). El Niño-related thermal stress coupled with upwelling-related ocean acidification negatively impacts cellular to population-level responses in pteropods along the California Current System with implications for increased bioenergetic costs. *Frontiers in Marine Science*, 5, 486. <https://doi.org/10.3389/fmars.2018.00486> [OO][WC]
- Bednaršek, N., Feely, R. A., Howes, E. L., Hunt, B. P. V., Kessouri, F., León, P., et al. (2019). Systematic review and meta-analysis toward synthesis of thresholds of ocean acidification impacts on calcifying pteropods and interactions with warming. *Frontiers in Marine Science*, 6, 227. <https://doi.org/10.3389/fmars.2019.00227> [WC]
- Bednaršek, N., Feely, R. A., Beck, M. W., Alin, S. R., Siedlecki, S. A., Calosi, P., et al. (2020). Exoskeleton dissolution with mechanoreceptor damage in larval Dungeness crab related to severity of present-day ocean acidification vertical gradients. *Science of the Total Environment*, 716, 136610. <https://doi.org/10.1016/j.scitotenv.2020.136610> [WC]
- Bednaršek, N., Carter, B. R., McCabe, R. M., Feely, R. A., Chavez, F. P., Elliott, M., et al. (in review). Rapid and persistent pelagic gastropods collapse due to extreme climatic events and ocean acidification. Submitted to *Nature Communications*. [OO]
- Beletsky, D., & Schwab, D. (2008). Climatological circulation in Lake Michigan. *Geophysical Research Letters*, 35(21), L21604. <https://doi.org/10.1029/2008GL035773> [GL]
- Bennett, N. J. (2019). Marine social science for the peopled seas. *Coastal Management*, 47(2), 244–253. <https://doi.org/10.1080/08920753.2019.1564958> [PAC]
- Bennett, N. J., Blythe, J., Tyler, S., & Ban, N. C. (2016). Communities and change in the anthropocene: Understanding social-ecological vulnerability and planning adaptations to multiple interacting exposures. *Regional Environmental Change*, 16(4), 907–926. [WC]
- Bennington, V., McKinley, G. A., Kimura, N., & Wu, C. H. (2010). General circulation of Lake Superior: Mean, variability, and trends from 1979 to 2006. *Journal of Geophysical Research: Oceans*, 115(C12), C12015. <https://doi.org/10.1029/2010JC006261> [GL]
- Bennington, V., McKinley, G. A., Urban, N. R., & McDonald, C. P. (2012). Can spatial heterogeneity explain the perceived imbalance in Lake Superior's carbon budget? A model study. *Journal of Geophysical Research: Biogeosciences*, 117(G3), G03020. <https://doi.org/10.1029/2011JG001895> [GL]
- Benthuisen, J., Thomas, L. N., & Lentz, S. J. (2015). Rapid generation of upwelling at a shelf break caused by buoyancy shutdown. *Journal of Physical Oceanography*, 45(1), 294–312. [MAB]
- Bercel, T. L., & Kranz, S. A. (2019). Insights into carbon acquisition and photosynthesis in *Karenia brevis* under a range of CO<sub>2</sub> concentrations. *Progress in Oceanography*, 172, 65–76. <https://doi.org/10.1016/j.pocean.2019.01.011> [SAG]
- Berman, M., & Schmidt, J. I. (2019). Economic effects of climate change in Alaska. *Weather, Climate, and Society*, 11, 245–258. <https://doi.org/10.1175/WCAS-D-18-0056.1> [ARC]
- Bignami, S., Enochs, I. C., Manzello, D. P., Sponaugle, S., & Cowen, R. K. (2013). Ocean acidification alters the otoliths of a pantropical fish species with implications for sensory function. *Proceedings of the National Academy of Sciences USA*, 110(18), 7366–7370. <https://doi.org/10.1073/pnas.1301365110> [FLC]
- Bishop, R. C., Chapman, D. J., Kanninen, B. J., Krosnick, J. A., Leeworthy, B., & Meade, N. F. (2011). Total economic value for protecting and restoring Hawaiian coral reef ecosystems: Final report. NOAA Technical Memorandum CRCP 16. NOAA Office of National Marine Sanctuaries, Office of Response and Restoration, and Coral Reef Conservation Program. 406 pp. [PAC]

- Bluhm, B. A., Iken, K., Mincks, S. L., Sirneko, B. I., & Holladay, B. A. (2009). Community structure of epibenthic megafauna in the Chukchi Sea. *Aquatic Biology*, 7, 269–293. <https://doi.org/10.3354/ab00198> [ARC]
- Boehme, S. E., Sabine, C. L., & Reimers, C. E. (1998). CO<sub>2</sub> fluxes from a coastal transect: A time-series approach. *Marine Chemistry*, 63(1-2), 49–67. [MAB]
- Boulais, M., Chenevert, K. J., Demey, A. T., Darrow, E. S., Robison, M. R., Roberts, J. P., & Volety, A. (2017). Oyster reproduction is compromised by acidification experienced seasonally in coastal regions. *Scientific Reports*, 7(1), 13276. <https://doi.org/10.1038/s41598-017-13480-3> [MAB]
- Boyd, P. W., Claustre, H., Levy, M., Siegel, D. A., & Weber, T. (2019). Multi-faceted particle pumps drive carbon sequestration in the ocean. *Nature*, 568, 327–335. <https://doi.org/10.1038/s41586-019-1098-2> [OO]
- Brainard, R. E., Oliver, T., McPhaden, M. J., Cohen, A., Venegas, R., Heenan, A., et al. (2018). Ecological impacts of the 2015/16 El Niño in the central equatorial Pacific. *Bulletin of the American Meteorological Society*, 99(1), S21–S26. <https://doi.org/10.1175/BAMS-D-17-0128.1> [PAC]
- Brander, L., & van Beukering, P. (2013). *The Total Economic Value of US Coral Reefs: A Review of the Literature*. Silver Spring, MD: NOAA Coral Reef Conservation Program, N/OCRM. [PAC]
- Breitburg, D. L., Salisbury, J., Bernhard, J. M., Cai, W.-J., Dupont, S., Doney, S. C., et al. (2015). And on top of all that... Coping with ocean acidification in the midst of many stressors. *Oceanography*, 28, 48–61. <https://doi.org/10.5670/oceanog.2015.31> [AK][ARC]
- Breslow, S. J., Sojka, B., Barnea, R., Basurto, X., Carothers, C., Charnley, S., et al. (2016). Conceptualizing and operationalizing human wellbeing for ecosystem assessment and management. *Environmental Science & Policy*, 66, 250–259. <https://doi.org/10.1016/j.envsci.2016.06.023> [WC]
- Bresnahan, P. J., Martz, T. R., Takeshita, Y., Johnson, K. S., & LaShomb, M. (2014). Best practices for autonomous measurement of seawater pH with the Honeywell Durafet. *Methods in Oceanography*, 9, 44–60. <https://doi.org/10.1016/J.MIO.2014.08.003> [WC]
- Bricker, S. B., Longstaff, B., Dennison, W., Jones, A., Boicourt, K., Wicks, C., & Woerner, J. (2008). Effects of nutrient enrichment in the nation's estuaries: A decade of change. *Harmful Algae*, 8(1), 21–32. [MAB]
- Brooke, S., Watts, M., Heil, A., Rhode, M., Mienis, F., Duineveld, G., et al. (2017). Distributions and habitat associations of deep-water corals in Norfolk and Baltimore Canyons, Mid-Atlantic Bight, USA. *Deep Sea Research Part II: Topical Studies in Oceanography*, 137, 131–147. <https://doi.org/10.1016/j.dsr2.2016.05.008> [MAB]
- Buckler, D. R., Mehrle, P. M., Cleveland, L., & Dwyer, F. J. (1987). Influence of pH on the toxicity of aluminium and other inorganic contaminants to East Coast striped bass. *Water, Air, and Soil Pollution*, 35(1-2), 97–106. [GL]
- Buitenhuis, E. T., Le Quéré, C., Bednaršek, N., & Schiebel, R. (2019). Large contribution of pteropods to shallow CaCO<sub>3</sub> export. *Global Biogeochemical Cycles*, 33, 458–468. <https://doi.org/10.1029/2018GB006110> [OO]
- Busch, D. S., & McElhany, P. (2016). Estimates of the direct effect of seawater pH on the survival rate of species groups in the California Current Ecosystem. *PLoS ONE*, 11(8), e0160669. <https://doi.org/10.1371/journal.pone.0160669> [WC]
- Busch, D. S., & McElhany, P. (2017). Using mineralogy and higher-level taxonomy as indicators of species sensitivity to pH: A case-study of Puget Sound. *Elementa*, 5, 53. <https://doi.org/10.1525/elementa.245> [WC]
- Busch, D. S., Harvey, C. J., & McElhany, P. (2013). Potential impacts of ocean acidification on the Puget Sound food web. *ICES Journal of Marine Science*, 70(4), 823–833. <https://doi.org/10.1093/icesjms/fst061> [INTRO][WC]
- Busch, D. S., Maher, M., Thibodeau, P., & McElhany, P. (2014). Shell condition and survival of Puget Sound pteropods are impaired by ocean acidification conditions. *PLoS ONE*, 9(8), e105884. <https://doi.org/10.1371/journal.pone.0105884> [INTRO][WC]
- Busch, D. S., Bennett-Mintz, J., Armstrong, C. T., Jewett, L., Gledhill, D., & Ombres, E. (2018). NOAA Ocean Acidification Program: Taking Stock and Looking Forward, A summary of the 2017 Principal Investigator's Meeting. U.S. Department of Commerce, NOAA Technical



- Memorandum OAR-OAP-1, 52 pp. [INTRO]
- Bushinsky, S. M., Takeshita, Y., & Williams, N. L. (2019). Observing changes in ocean carbonate chemistry: Our autonomous future. *Current Climate Change Reports*, 5, 207–220. <https://doi.org/10.1007/s40641-019-00129-8> [OO]
- Byrne, R. H., Mecking, S., Feely, R. A., & Liu, X. (2010). Direct observations of basin-wide acidification of the North Pacific Ocean. *Geophysical Research Letters*, 37, L02601. <https://doi.org/10.1029/2009GL040999> [OO]
- CAFF (2013). Arctic Biodiversity Assessment. Status and Trends in Arctic Biodiversity. Conservation of Arctic Flora and Fauna, Akureyri. Retrieved from <https://www.caff.is/assessment-series/arctic-biodiversity-assessment> [ARC]
- Cai, W.-J., Hu, X., Huang, W.-J., Murrell, M. C., Lehner, J. C., Lohrenz, S. E., et al. (2011). Acidification of subsurface coastal waters enhanced by eutrophication. *Nature Geoscience*, 4(11), 766–770. <https://doi.org/10.1038/ngeo1297> [SAG][MAB]
- Caldeira, K., & Wickett, M. E. (2003). Anthropogenic carbon and ocean pH. *Nature*, 425, 365. [INTRO]
- Calvo, L. (2018). *New Jersey Shellfish Aquaculture Situation and Outlook Report 2016 Production Year*, New Jersey Sea Grant Publication #18-931. Retrieved (September 2019) from <http://njseagrant.org/new-jersey-shellfish-aquaculture-situation-outlook-report-new/> [MAB]
- Carmack, E. C., Yamamoto-Kawai, M., Haine, T. W. N., Baron, S., Bluhm, B. A., Lique, C., et al. (2016). Freshwater and its role in the Arctic Marine System: Sources, disposition, storage, export, and physical and biogeochemical consequences in the Arctic and global oceans. *Journal of Geophysical Research—Biogeosciences*, 121(3), 675–717. <https://doi.org/10.1002/2015JG003140> [ARC]
- Caron, D. A., & Hutchins, D. A. (2012). The effects of changing climate on microzooplankton grazing and community structure: Drivers, predictions and knowledge gaps. *Journal of Plankton Research*, 35(2), 235–252. <https://doi.org/10.1093/plankt/fbs091> [SAG]
- Carter, B. R., Frölicher, T. L., Dunne, J. P., Rodgers, K. B., Slater, R. D., & Sarmiento, J. L. (2016). When can ocean acidification impacts be detected from decadal alkalinity measurements?. *Global Biogeochemical Cycles*, 30, 595–612. <https://doi.org/10.1002/2015GB005308> [OO]
- Carter, B. R., Feely, R. A., Mecking, S., Cross, J. N., Macdonald, A. M., Siedlecki, S. A., et al. (2017). Two decades of Pacific anthropogenic carbon storage and OA along Global Ocean Ship-based Hydrographic Investigations Program sections P16 and P02. *Global Biogeochemical Cycles*, 31(2), 306–327. <https://doi.org/10.1002/2016GB005485> [OO][AK]
- Carter, B. R., Feely, R. A., Wanninkhof, R., Kouketsu, S., Sonnerup, R. E., Pardo, P. C., et al. (2019a). Pacific anthropogenic carbon between 1991 and 2017. *Global Biogeochemical Cycles*, 33(5), 597–617. <https://doi.org/10.1029/2018GB006154> [OO][WC] [AK]
- Carter, B. R., Williams, N. L., Evans, W., Fassbender, A. J., Barbero, L., Hauri, C., et al. (2019b). Time of detection as a metric for prioritizing between climate observation quality, frequency, and duration. *Geophysical Research Letters*, 46(7), 3853–3861. <https://doi.org/10.1029/2018GL080773> [WC]
- Chambers, R. C., Candelmo, A., Habeck, E., Poach, M., Wieczorek, D., Cooper, K., et al. (2014). Effects of elevated CO<sub>2</sub> in the early life stages of summer flounder, *Paralichthys dentatus*, and potential consequences of ocean acidification. *Biogeosciences*, 11(6), 1613–1626. [MAB][NE]
- Chan, F., Barth, J. A., Blanchette, C. A., Byrne, R. H., Chavez, F., Cheriton, O., et al. (2017). Persistent spatial structuring of coastal ocean acidification in the California Current System. *Scientific Reports*, 7, 2526. <https://doi.org/10.1038/s41598-017-02777-y> [WC]
- Chapra, S. C., Dove, A., & Warren, G. J. (2012). Long-term trends of Great Lakes major ion chemistry. *Journal of Great Lakes Research*, 38(3), 550–560. [GL]
- Chavez, F. P., Pennington, J. T., Michisaki, R. P., Blum, M., Chavez, G. M., Friederich, J., et al. (2017) Climate variability and change: Response of a coastal ocean ecosystem. *Oceanography*, 30(4), 128–145. <https://doi.org/10.5670/oceanog.2017.429> [WC]
- Chen, C.-T. A., Wei, C.-L., & Rodman, M. R. (1985). Carbonate chemistry of the Bering Sea. Report no. DOE/EV/10611-5; Contract no.: DOE-AT06-81EV10611. Washington, D.C.: U.S. Department of Energy, Office of Energy Research, Office of

- Basic Energy Sciences, Carbon Dioxide Research Division. [ARC]
- Chen, S., Hu, C., Barnes, B. B., Wanninkhof, R., Cai, W.-J., Barbero, L., & Pierrot, D. (2019). A machine learning approach to estimate surface ocean  $p\text{CO}_2$  from satellite measurements. *Remote Sensing of Environment*, 228, 203–226. <https://doi.org/10.1016/j.rse.2019.04.019> [SAG]
- Chesapeake Bay Foundation (2020). The State of the Bay's Oyster Fishery. Retrieved from <https://www.cbf.org/issues/fisheries/the-state-of-todays-oyster-fishery.html>. [MAB]
- CITES (2003). Progress on the implementation of the review of significant trade (phases IV and V). Report to the Nineteenth Meetings of the CITES Animals Committee. AC19 Doc. 8.3. Convention on International Trade in Endangered Species [FLC]
- Clark, H. R., & Gobler, C. J. (2016). Diurnal fluctuations in  $\text{CO}_2$  and dissolved oxygen concentrations do not provide a refuge from hypoxia and acidification for early-life-stage bivalves. *Marine Ecology Progress Series*, 558, 1–14. <https://doi.org/10.3354/meps11852> [MAB]
- Claudi, R., Graves, A., Taraborelli, A. C., Prescott, R. J., & Mastitsky, S. E. (2012). Impact of pH on survival and settlement of dreissenid mussels. *Aquatic Invasions*, 7(2), 21–28. <https://doi.org/10.3391/ai.2012.7.1.003> [GL]
- Clements, J. C., & Chopin, T. (2017). Ocean acidification and marine aquaculture in North America: Potential impacts and mitigation strategies. *Reviews in Aquaculture*, 9(4), 326–341. [MAB] [NE]
- Clements, J. C., & Hunt, H. L. (2014). Influence of sediment acidification and water flow on sediment acceptance and dispersal of juvenile soft-shell clams (*Mya arenaria* L.). *Journal of Experimental Marine Biology and Ecology*, 453, 62–69. [MAB][NE]
- Clements, J. C., & Hunt, H. L. (2015). Marine animal behaviour in a high  $\text{CO}_2$  ocean. *Marine Ecology Progress Series*, 536, 259–279. <https://doi.org/10.3354/meps11426> [AK]
- Clements, J. C., & Hunt, H. L. (2018). Testing for sediment acidification effects on within-season variability in juvenile soft-shell clam (*Mya arenaria*) abundance on the northern shore of the Bay of Fundy. *Estuaries and Coasts*, 41(2), 471–483. [INTRO][NE]
- Coffey, W. D., Nardone, J. A., Yarram, A., Long, W. C., Swiney, K. M., Foy, R. J., & Dickinson, G. H. (2017). Ocean acidification leads to altered micromechanical properties of the mineralized cuticle in juvenile red and blue king crabs. *Journal of Experimental Marine Biology and Ecology*, 495, 1–12. <https://doi.org/10.1016/j.jembe.2017.05.011> [AK]
- Colburn, L. L., Jepson, M., Weng, C., Seara, T., Weiss, J., & Hare, J. A. (2016). Indicators of climate change and social vulnerability in fishing dependent communities along the Eastern and Gulf Coasts of the United States. *Marine Policy*, 74, 323–333. [NE]
- Cooley, S. R., & Doney, S. C. (2009). Anticipating ocean acidification's economic consequences for commercial fisheries. *Environmental Research Letters*, 4, 024007. <https://doi.org/10.1088/1748-9326/4/2/024007> [OO] [PAC]
- Cooley, S. R., Lucey, N., Kite-Powell, H. L., & Doney, S. C. (2012). Nutrition and income from molluscs today imply vulnerability to ocean acidification tomorrow. *Fish and Fisheries*, 13, 182–215. <https://doi.org/10.1111/j.1467-2979.2011.00424.x> [OO]
- Cooley, S. R., Rheuban, J., Hart, D., Luu, V., Glover, D., Hare, J., & Doney, S. (2015). An integrated assessment model for helping the United States sea scallop (*Placopecten magellanicus*) fishery plan ahead for ocean acidification and warming. *PLoS ONE*, 10(5), e0124145. <https://doi.org/10.1371/journal.pone.0124145> [NE]
- Cooley, S. R., Ono, C. R., Melcer, S., & Roberson, J. (2016). Community-level actions that can address ocean acidification. *Frontiers in Marine Science*, 2, 128. <https://doi.org/10.3389/fmars.2015.00128> [OO]
- Cooper, H. L., Potts, D. C., & Paytan, A. (2016). Effects of elevated  $p\text{CO}_2$  on the survival, growth, and moulting of the Pacific krill species, *Euphausia pacifica*. *ICES Journal of Marine Science*, 74(4), 1005–1012. <https://doi.org/10.1093/icesjms/fsw021> [ARC]
- Crane, K., & Ostrovskiy, A. (2015). Russian-American Long-Term Census of the Arctic: RUSALCA. *Oceanography*, 28(3), 18–23. <https://doi.org/10.5670/oceanog.2015.54> [ARC]
- Crook, E. D., Cohen, A. L., Rebolledo-Vieyra, M.,

- Hernandez, L., & Paytan, A. (2013). Reduced calcification and lack of acclimatization by coral colonies growing in areas of persistent natural acidification. *Proceedings of the National Academy of Sciences USA*, *110*, 11044–11049. <https://doi.org/10.1073/pnas.1301589110> [FLC]
- Cross, J. N., Mathis, J. T., Bates, N. R., & Byrne, R. H. (2013). Conservative and non-conservative variations of total alkalinity on the southeastern Bering Sea shelf. *Marine Chemistry*, *154*, 100–112. <https://doi.org/10.1016/j.marchem.2013.05.012> [AK]
- Cross, J. N., Mathis, J. T., Lomas, M. W., Moran, S. B., Baumann, M. S., Shull, D. H., et al. (2014). Integrated assessment of the carbon budget in the southeastern Bering Sea. *Deep Sea Research Part II*, *109*, 112–124. <https://doi.org/10.1016/j.dsr2.2014.03.003> [AK]
- Cross, J. N., Mordy, C. W., Tabisola, H. M., Meining, C., Cokelet, E. D., & Stabeno, P. J. (2016). Innovative technology development for Arctic Exploration. *Oceans 2015*, MTS-IEEE Washington, D.C. <https://doi.org/10.23919/OCEANS.2015.7404632> [ARC]
- Cross, J. N., Mathis, J. T., Pickart, R. S., & Bates, N. R. (2018). Formation and transport of corrosive water in the Pacific Arctic region. *Deep-Sea Research Part II*, *152*, 67–81. <https://doi.org/10.1016/j.dsr2.2018.05.020> [ARC]
- Cyronak, T., Santos, I. R., & Eyre, B. D. (2013). Permeable coral reef sediment dissolution driven by elevated  $p\text{CO}_2$  and pore water advection. *Geophysical Research Letters*, *40*(18), 4876–4881. <https://doi.org/10.1002/grl.50948> [FLC]
- Cyronak, T., Andersson, A. J., Langdon, C., Albright, R., Bates, N. R., Caldeira, K., et al. (2018). Taking the metabolic pulse of the world's coral reefs. *PLoS ONE*, *13*(1), e0190872. <https://doi.org/10.1371/journal.pone.0190872> [FLC]
- Darnis, G., Barber, D. G., & Fortier, L. (2008). Sea ice and the onshore-offshore gradient in pre-winter zooplankton assemblages in southeastern Beaufort Sea. *Journal of Marine Systems*, *74*(3-4), 994–1011. <https://doi.org/10.1016/j.jmarsys.2007.09.003> [ARC]
- Davis, C. V., Rivest, E. B., Hill, T. M., Gaylord, B., Russell, A. D., & Sanford, E. (2017). Ocean acidification compromises a planktic calcifier with implications for global carbon cycling. *Scientific Reports*, *7*(1), 2225. <https://doi.org/10.1038/s41598-017-01530-9> [WC]
- DeCarlo, T. M., Cohen, A. L., Barkley, H. C., Cobban, Q., Young, C., Shamberger, K. E., et al. (2015). Coral macrobioerosion is accelerated by OA and nutrients. *Geology*, *43*(1), 7–10. <https://doi.org/10.1130/G36147.1> [PAC]
- de Moel, H., Ganssen, G. M., Peeters, F. J. C., Jung, S. J. A., Kroon, D., Brummer, G. J. A., & Zeebe, R. E. (2009). Planktic foraminiferal shell thinning in the Arabian Sea due to anthropogenic ocean acidification? *Biogeosciences*, *6*, 1917–1925. [INTRO]
- Dery, A., Collard, M., & Dubois, P. (2017). Ocean acidification reduces spine mechanical strength in euechinoid but not in cidaroid sea urchins. *Environmental Science & Technology*, *51*(7), 3640–3648. <https://doi.org/10.1021/acs.est.6b05138> [FLC]
- De Stasio, B. T., Hill, D. K., Kleinhans, J. M., Nibbelink, N. P., & Magnuson, J. J. (1996). Potential effects of global climate change on small north-temperate lakes: Physics, fish, and plankton. *Limnology and Oceanography*, *41*(5), 1136–1149. [GL]
- DeVries, T. (2014). The oceanic anthropogenic  $\text{CO}_2$  sink: Storage, air-sea fluxes, and transports over the industrial era. *Global Biogeochemical Cycles*, *28*, 631–647. <https://doi.org/10.1002/2013GB004739> [OO]
- DeVries, T., Holzer, M., & Primeau, F. (2017). Recent increase in oceanic carbon uptake driven by weaker upper-ocean overturning. *Nature*, *542*(7640), 215–218. <https://doi.org/10.1038/nature21068> [OO]
- De Wit, P., Dupont, S., & Thor, P. (2016). Selection on oxidative phosphorylation and ribosomal structure as a multigenerational response to ocean acidification in the common copepod *Pseudocalanus acuspes*. *Evolutionary Applications*, *9*, 1112–1123. <https://doi.org/10.1111/eva.12335> [ARC]
- Di Lorenzo, E., & Mantua, N. (2016). Multi-year persistence of the 2014/15 North Pacific marine heatwave. *Nature Climate Change*, *6*, 1042–1047. <https://doi.org/10.1038/nclimate3082> [OO]
- Dixon, G. B., Davies, S. W., Aglyamova, G. V., Meyer, E., Bay, L. K., & Matz, M. V. (2015). Genomic determinants of coral heat tolerance across latitudes. *Science*, *348*(6242), 1460–1462.



<https://doi.org/10.1126/science.1261224> [FLC]

- Dore, J. E., Lukas, R., Sadler, D. W., Church, M. J., & Karl, D. M. (2009). Physical and biogeochemical modulation of ocean acidification in the central North Pacific. *Proceedings of the National Academy of Sciences of the United States of America*, 106(30), 12235–12240. <https://doi.org/10.1073/pnas.0906044106> [PAC]
- Duarte, C. M., Hendriks, I. E., Moore, T. S., Olsen, Y. S., & Steckbauer, A. (2013). Is ocean acidification an open-ocean syndrome? Understanding anthropogenic impacts on seawater pH. *Estuaries and Coasts*, 36, 221–236. <https://doi.org/10.1007/s12237-013-9594-3> [FLC]
- Dubik, B. A., Clark, E. C., Young, T., Zigler, S. B. J., Provost, M. M., Pinsky, M. L., & Martin, K. S. (2019). Governing fisheries in the face of change: Social responses to long-term geographic shifts in a US fishery. *Marine Policy*, 99, 243–251. [MAB]
- Duncan, B. E., Higgason, K. D., Suchanek, T. H., Largier, J., Stachowicz, J., Allen, S., et al. (2014). *Ocean Climate Indicators: A Monitoring Inventory and Plan for Tracking Climate Change in the North-central California Coast and Ocean Region*. Report of a Working Group of the Gulf of the Farallones National Marine Sanctuary Advisory Council. Marine Sanctuaries Conservation Series ONMS-14-09. Silver Spring, MD: U.S. Department of Commerce, National Oceanic and Atmospheric Administration, Office of National Marine Sanctuaries. 81 pp. [WC]
- Dunne, J. P., John, J. G., Adcroft, A. J., Griffies, S. M., Hallberg, R. W., Shevliakova, E., et al. (2012a). GFDL's ESM2 global coupled climate-carbon earth system models. Part I: Physical formulation and baseline simulation characteristics. *Journal of Climate*, 25(19), 6646–6665. <https://doi.org/10.1175/JCLI-D-11-00560.1> [OO]
- Dunne, J. P., Hales, B., & Toggweiler, J. R. (2012b). Global calcite cycling constrained by sediment preservation controls. *Global Biogeochemical Cycles*, 26(3), GB3023. <https://doi.org/10.1029/2010GB003935> [OO]
- Dunne, J. P., John, J. G., Shevliakova, E., Stouffer, R. J., Krasting, J. P., Malyshev, S. L., et al. (2013). GFDL's ESM2 global coupled climate-carbon earth system models. Part II: Carbon system formulation and baseline simulation characteristics. *Journal of Climate*, 26(7), 2247–2267. <https://doi.org/10.1175/JCLI-D-12-00150.1> [OO]
- Edwards, P. E. T. (Ed.) (2013). *Summary Report: The Economic Value of U.S. Coral Reefs*. Silver Spring, MD: NOAA Coral Reef Conservation Program. 28 pp. [NAT]
- Ekstrom, J. A., Suatoni, L., Cooley, S. R., Pendleton, L. H., Waldbusser, G. G., Cinner, J. E., et al. (2015). Vulnerability and adaptation of US shellfisheries to ocean acidification. *Nature Climate Change*, 5, 207–214. <https://doi.org/10.1038/nclimate2508> [SAG][MAB][NE]
- Engström-Öst, J., Glippa, O., Feely, R. A., Kanerava, M., Keister, J. E., Alin, S. R., et al. (2019). Eco-physiological responses of copepods and pteropods to ocean warming and acidification. *Scientific Reports*, 9(1), 4748. <https://doi.org/10.1038/s41598-019-41213-1> [OO][WC]
- Enochs, I. C., Manzello, D. P., Carlton, R., Schopmeyer, S., van Hooidek, R., & Lirman, D. (2014). Effects of light and elevated  $p\text{CO}_2$  on the growth and photochemical efficiency of *Acropora cervicornis*. *Coral Reefs*, 33, 477–485. <https://doi.org/10.1007/s00338-014-1132-7> [FLC]
- Enochs, I. C., Manzello, D. P., Donham, E. M., Kolodziej, G., Okano, R., Johnston, L., et al. (2015a). Shift from coral to macroalgae dominance on a volcanically acidified reef. *Nature Climate Change*, 5, 1083–1089. <https://doi.org/10.1038/nclimate2758> [INTRO][FLC]
- Enochs, I. C., Manzello, D. P., Carlton, R. D., Graham, D. M., Ruzicka, R., & Colella, M. A. (2015b). Ocean acidification enhances the bioerosion of a common coral reef sponge: implications for the persistence of the Florida Reef Tract. *Bulletin of Marine Science*, 92(2), 271–290. [FLC]
- Enochs, I. C., Manzello, D. P., Kolodziej, G., Noonan, S. H. C., Valentino, L., & Fabricius, K. E. (2016a). Enhanced macroboring and depressed calcification drive net dissolution at high- $\text{CO}_2$  coral reefs. *Proceedings of the Royal Society B: Biological Sciences*, 283(1842), 20161742. <https://doi.org/10.1098/rspb.2016.1742> [PAC][FLC]
- Enochs, I. C., Manzello, D. P., Wirshing, H. H., Carlton, R., Serafy, J. (2016b). Micro-CT analysis of the Caribbean octocoral *Eunicea flexuosa* subjected to elevated  $p\text{CO}_2$ . *ICES Journal of Marine Science*, 73(3), 910–919. <https://doi.org/10.1093/icesjms/fsv159> [FLC]
- Enochs, I. C., Manzello, D. P., Jones, P. R., Stamates, S. J., & Carsey, T. P. (2019). Seasonal carbon-

- ate chemistry dynamics on southeast Florida coral reefs: Localized acidification hotspots from navigational inlets. *Frontiers in Marine Science*, 6, 160. <https://doi.org/10.3389/fmars.2019.00160> [FLC][SAG]
- Errera, R. M., Yvon-Lewis, S., Kessler, J. D., & Campbell, L. (2014). Responses of the dinoflagellate *Karenia brevis* to climate change: pCO<sub>2</sub> and sea surface temperatures. *Harmful Algae* 37, 110–116. <https://doi.org/10.1016/j.hal.2014.05.012> [SAG]
- Evans, L. S., Hicks, C. C., Fidelman, P., Tobin, R. C., & Perry, A. L. (2013). Future scenarios as a research tool: Investigating climate change impacts, adaptation options and outcomes for the Great Barrier Reef, Australia. *Human Ecology*, 41(6), 841–857. <https://doi.org/10.1007/s10745-013-9601-0> [WC]
- Evans, W., Mathis, J. T., Cross, J. N., Bates, N. R., Frey, K. E., Else, B. G. T., et al. (2015). Sea-air CO<sub>2</sub> exchange in the western Arctic coastal ocean. *Global Biogeochemical Cycles*, 29(8), 1190–1209. <https://doi.org/10.1002/2015GB005153> [ARC]
- Evans, W., Pocock, K., Hare, A., Weekes, C., Hales, B., Jackson, J., et al. (2019). Marine CO<sub>2</sub> patterns in the northern Salish Sea. *Frontiers in Marine Science*, 5, 536. <https://doi.org/10.3389/fmars.2018.00536> [WC]
- Eyre, B. D., Andersson A. J., & Cyronak T. (2014). Benthic coral reef calcium carbonate dissolution in an acidifying ocean. *Nature Climate Change*, 4, 969–976 [FLC]
- Eyre, B. D., Cyronak, T., Drupp, P., De Carlo, E. H., Sachs, J. P., & Andersson, A. J. (2018). Coral reefs will transition to net dissolving before end of century. *Science*, 359, 908–911. [FLC]
- Eyring, V., Bony, S., Meehl, G. A., Senior, C. A., Stevens, B., Stouffer, R. J., & Taylor, K. E. (2016). Overview of the Coupled Model Intercomparison Project Phase 6 (CMIP6) experimental design and organization. *Geoscientific Model Development*, 9, 1937–1958. [OO]
- Fabricius, K. E., Langdon, C., Uthicke, S., Humphrey, C., Noonan, S., De'ath, G., et al. (2011). Losers and winners in coral reefs acclimatized to elevated carbon dioxide concentrations. *Nature Climate Change*, 1, 165–169. <https://doi.org/10.1038/nclimate1122> [FLC]
- Fabry, V. J., Seibel, B. A., Feely, R. A., & Orr, J. C. (2008). Impacts of ocean acidification on marine fauna and ecosystem processes. *ICES Journal of Marine Science*, 65(3), 414–432. <https://doi.org/10.1093/icesjms/fsn048> [INTRO]
- Fabry, V. J., McClintock, J. B., Mathis, J. T., & Grebmeier, J. M. (2009). Ocean acidification at high latitudes: the bellweather. *Oceanography*, 22(4), 160–171. [AK]
- Fall, J. A. (2012). *Subsistence in Alaska – A Year 2010 Update*. Anchorage, AK: Division of Subsistence, Alaska Department of Fish and Game. [AK]
- Fay, G., Link, J. S., & Hare, J. A. (2017). Assessing the effects of ocean acidification in the northeast US using an end-to-end marine ecosystem model. *Ecological Modelling*, 347, 1–10. <https://doi.org/10.1016/j.ecolmodel.2016.12.016> [NE]
- Feely, R. A., Sabine, C. L., Lee, K., Berelson, W., Kleypas, J., Fabry, V. J., & Millero, F. J. (2004). Impact of anthropogenic CO<sub>2</sub> on the CaCO<sub>3</sub> system in the oceans. *Science*, 305(5682), 362–366. <https://doi.org/10.1126/science.1097329> [INTRO][OO]
- Feely, R. A., Takahashi, T., Wanninkhof, R., McPhaden, M. J., Cosca, C. E., Sutherland, S. C., & Carr, M.-E. (2006). Decadal variability of the air-sea CO<sub>2</sub> fluxes in the equatorial Pacific Ocean. *Journal of Geophysical Research*, 111(C08), C08S90. <https://doi.org/10.1029/2005JC003129> [NAT]
- Feely, R. A., Sabine, C. L., Hernandez-Ayon, J. M., Ianson, D., & Hales, B. (2008). Evidence for upwelling of corrosive “acidified” water onto the Continental Shelf. *Science*, 320(5882), 1490–1492. <https://doi.org/10.1126/science.1155676> [INTRO][WC]
- Feely, R. A., Doney, S. C., & Cooley, S. R. (2009). Ocean acidification: Present conditions and future changes in a high-CO<sub>2</sub> world. *Oceanography*, 22(4), 36–47. <https://doi.org/10.5670/oceanog.2009.95> [INTRO][NAT][OO]
- Feely, R. A., Alin, S. R., Newton, J., Sabine, C. L., Warner, M., Devol, A., et al. (2010). The combined effects of ocean acidification, mixing, and respiration on pH and carbonate saturation in an urbanized estuary. *Estuarine, Coastal and Shelf Science*, 88(4), 442–449. [WC]
- Feely, R., Sabine, C., Byrne, R., Millero, F., Dickson, A., Wanninkhof, R., et al. (2012). Decadal

- changes in the aragonite and calcite saturation state of the Pacific Ocean. *Global Biogeochemical Cycles*, 26(3), GB3001. <https://doi.org/10.1029/2011GB004157> [OO][WC]
- Feely, R. A., Alin, S. R., Carter, B., Bednaršek, N., Hales, B., Chan, F., et al. (2016). Chemical and biological impacts of OA along the west coast of North America. *Estuarine, Coastal and Shelf Science*, 183(A), 260–270. <https://doi.org/10.1016/j.ecss.2016.08.043> [OO][WC]
- Feely, R. A., Okazaki, R. R., Cai, W.-J., Bednaršek, N., Alin, S. R., Byrne, R. H., & Fassbender, A. (2018). The combined effects of acidification and hypoxia on pH and aragonite saturation in the coastal waters of the California Current Ecosystem and the northern Gulf of Mexico. *Continental Shelf Research*, 152, 50–60. <https://doi.org/10.1016/j.csr.2017.11.002> [WC][SAG]
- Fennel, K., Alin, S., Barbero, L., Evans, W., Bourgeois, T., Cooley, S., et al. (2019). Carbon cycling in the North American coastal ocean: a synthesis. *Biogeosciences*, 16(6), 1281–1304. <https://doi.org/10.5194/bg-16-1281-2019> [SAG]
- Finkelstein, S. A., Bunbury, J., Gajewski, K., Wolfe, A. P., Adams, J. K., & Devlin, J. E. (2014). Evaluating diatom-derived Holocene pH reconstructions for Arctic lakes using an expanded 171-lake training set. *Journal of Quaternary Science*, 29(3), 249–260. [GL]
- Fissel, B., Dalton, M., Garber-Yonts, B., Haynie, A., Kasperski, S., Lee, J., et al. (2017). *Stock Assessment and Fishery Evaluation Report for the Groundfish Fisheries of the Gulf of Alaska and Bering Sea/Aleutian Islands Area: Economic Status of the Groundfish Fisheries Off Alaska, 2016*. Seattle, WA: Economic and Social Sciences Research Program, Resource Ecology and Fisheries Management Division, Alaska Fisheries Science Center, National Marine Fisheries Service, National Oceanic and Atmospheric Administration. 425 pp. [AK]
- Gaines, S. D., Costello, C., Owashi, B., Mangin, T., Bone, J., Molinos, J. G., et al. (2018). Improved fisheries management could offset many negative effects of climate change. *Science Advances*, 4(8), eaao1378. <https://doi.org/10.1126/sciadv.aao1378> [AK]
- Garcia, A. C., Bargu, S., Dash, P., Rabalais, N. N., Sutor, M., Morrison, W., & Walker, N. D. (2010). Evaluating the potential risk of microcystins to blue crab (*Callinectes sapidus*) fisheries and human health in a eutrophic estuary. *Harmful Algae* 9(2), 134–143. <https://doi.org/10.1016/j.hal.2009.08.011> [SAG]
- García-Reyes, M., Sydeman, W. J., Schoeman, D. S., Rykaczewski, R. R., Black, B. A., Smit, A. J., & Bograd, S. J. (2015). Under pressure: Climate change, upwelling, and eastern boundary upwelling ecosystems. *Frontiers in Marine Science*, 2, 109. doi:10.3389/fmars.2015.00109 [WC]
- Gardner, T. A., Côté, I. M., Gill, J. A., Grant, A., & Watkinson, A. R. (2003). Long-term region-wide declines in Caribbean corals. *Science*, 301(5635), 958–960. <https://doi.org/10.1126/science.1086050> [FLC]
- Gates, J. M. (2009). Investing in our future: The economic case for rebuilding Mid-Atlantic fish populations: Pew Environment Group. [MAB]
- Gattuso, J.-P., Magnan, A., Bille, R., Cheung, W. W. L., Howes, E. L., Joos, F., et al. (2015). Contrasting futures for ocean and society from different anthropogenic CO<sub>2</sub> emission scenarios. *Science*, 349(6243), aac4722. <https://doi.org/10.1126/science.aac4722> [INTRO] [OO]
- Ge, C., Chai, Y., Wang, H., & Kan, M. (2017). Ocean acidification: One potential driver of phosphorus eutrophication. *Marine Pollution Bulletin*, 115(1-2), 149–153. <https://doi.org/10.1016/j.marpolbul.2016.12.016> [NE]
- Georgian, S. E., DeLeo, D., Durkin, A., Gomez, C. E., Kurman, M., Lunden, J. J., & Cordes, E. E. (2016). Oceanographic patterns and carbonate chemistry in the vicinity of cold-water coral reefs in the Gulf of Mexico: Implications for resilience in a changing ocean. *Limnology and Oceanography*, 61, 648–665. <https://doi.org/10.1002/lno.10242> [SAG]
- Gibble, C., Duerr, R., Bodenstern, B., Lindquist, K., Lindsey, J., Beck, J., et al. (2018). Investigation of a largescale Common Murre (*Uria aalge*) mortality event in California, USA, in 2015. *Journal of Wildlife Diseases*, 54(3), 569–574. <https://doi.org/10.7589/2017-07-179> [WC]
- Gill, D., Rowe, M., & Joshi, S. J. (2018). Fishing in greener waters: Understanding the impact of harmful algal blooms on Lake Erie anglers and the potential for adoption of a forecast model. *Journal of Environmental Management*, 227, 248–255. <https://doi.org/10.1016/j.jenvman.2018.08.074> [GL]
- Giltz, S. M., & Taylor, C. M. (2017). Reduced growth



- and survival in the larval blue crab *Callinectes sapidus* under predicted ocean acidification. *Journal of Shellfish Research*, 36(2), 481–485. <https://doi.org/10.2983/035.036.0219> [MAB]
- Gingerich, P. D. (2019). Temporal scaling of carbon emission and accumulation rates: Modern anthropogenic emissions compared to estimates of PETM onset accumulation. *Paleoceanography and Paleoclimatology*, 34(3), 329–335. [INTRO]
- Glandon, H. L., & Miller, T. J. (2016). No effect of high  $p\text{CO}_2$  on juvenile blue crab, *Callinectes sapidus*, growth and consumption despite positive responses to concurrent warming. *ICES Journal of Marine Science*, 74(4), 1201–1209. <https://doi.org/10.1093/icesjms/fsw171> [MAB]
- Glandon, H. L., Kilbourne, K. H., Schijf, J., & Miller, T. J. (2018). Counteractive effects of increased temperature and  $p\text{CO}_2$  on the thickness and chemistry of the carapace of juvenile blue crab, *Callinectes sapidus*, from the Patuxent River, Chesapeake Bay. *Journal of Experimental Marine Biology and Ecology*, 498, 39–45. [MAB]
- Glaspie, C. N., Seitz, R. D., & Lipcius, R. N. (2017). The perfect storm: Extreme weather drives and predation maintains phase shift in dominant Chesapeake Bay bivalve. *bioRxiv*, 224097. [MAB]
- Gledhill, D. K., Wanninkhof, R., Millero, F. K., & Eakin, M. (2008). Ocean acidification of the Greater Caribbean region 1996–2006. *Journal of Geophysical Research—Oceans*, 113(10), C10031. <https://doi.org/10.1029/2007JC004629> [PAC]
- Gledhill, D. K., Wanninkhof, R., & Eakin, C. M. (2009). Observing ocean acidification from space. *Oceanography*, 22(4), 48–59. <https://doi.org/10.5670/oceanog.2009.96> [FLC]
- Gledhill, D. K., White, M. M., Salisbury, J., Thomas, H., Mlsna, I., Liebman, M., et al. (2015). Ocean and coastal acidification of New England and Nova Scotia. *Oceanography*, 28(2), 182–197. <https://doi.org/10.5670/oceanog.2015.41> [OE]
- Glenn, S., Arnone, R., Bergmann, T., Bissett, W. P., Crowley, M., Cullen, J., et al. (2004). Biogeochemical impact of summertime coastal upwelling on the New Jersey Shelf. *Journal of Geophysical Research: Oceans*, 109(C12), C12S02. <https://doi.org/10.1029/2003JC002265> [MAB]
- Gobler, C. J., & Baumann, H. (2016). Hypoxia and acidification in ocean ecosystems: coupled dynamics and effects on marine life. *Biology Letters*, 12(5), 20150976. [NE]
- Gobler, C. J., & Talmage, S. C. (2014). Physiological response and resilience of early life-stage Eastern oysters (*Crassostrea virginica*) to past, present and future ocean acidification. *Conservation Physiology*, 2(1), cou004. <https://doi.org/10.1093/conphys/cou004> [MAB][NE]
- Goethel, C. L., Grebmeier, J. M., Cooper, L. W., & Miller, T. J. (2017). Implications of ocean acidification in the Pacific Arctic: Experimental responses of three Arctic bivalves to decreased pH and food availability. *Deep Sea Research Part II*, 144, 112–124. <https://doi.org/10.1016/j.dsr2.2017.08.013> [ARC]
- Goldsmith, K. A., Lau, S., Poach, M. E., Sakowicz, G. P., Trice, T. M., Ono, C. R., et al. (2019). Scientific considerations for acidification monitoring in the U.S. Mid-Atlantic Region. *Estuarine, Coastal and Shelf Science*, 225, 106189. <https://doi.org/10.1016/j.ecss.2019.04.023> [MAB]
- Gorstein, M., Dillard, M., Loerzel, J., Edwards, P., & Levine, A. (2016). *National Coral Reef Monitoring Program Socioeconomic Monitoring Component: Summary Findings for South Florida, 2014*. U.S. Department of Commerce, NOAA Technical Memorandum, NOAA-TM-NOS-CRCP-25, 57 pp. + Appendices. <https://doi.org/10.7289/V5VH5KV5> [FL]
- Gorstein, M., Loerzel, J., Edwards, P., Levine, A., & Dillard, M. (2017). *National Coral Reef Monitoring Program Socioeconomic Monitoring Component: Summary Findings for Puerto Rico, 2015*. U.S. Department of Commerce, NOAA Technical Memorandum, NOAA-TM-NOS-CRCP-28, 64 pp. + Appendices. <https://doi.org/10.7289/V5BP00V9> [FL]
- Gorstein, M., Loerzel, J., Edwards, P., Levine, A., & Dillard, M. (2018a). *National Coral Reef Monitoring Program Socioeconomic Monitoring Component: Summary Findings for Guam, 2016*. U.S. Department of Commerce, NOAA Technical Memorandum, NOAA-TM-NOS-CRCP-32, 64 pp. + Appendices. <https://doi.org/10.25923/kpvd-mj07> [PAC]
- Gorstein, M., Loerzel, J., Levine, A., Edwards, P., & Dillard, M. (2018b). *National Coral Reef Mon-*

- itoring Program Socioeconomic Monitoring Component: Summary Findings for Hawai'i, 2015. U.S. Department of Commerce, NOAA Technical Memorandum, NOAA-TM-NOS-CRCP-30, 69 pp. + Appendices. [PAC]
- Gorstein, M., Loerzel, J., Edwards, P., Levine, A., & Dillard, M. (2019). *National Coral Reef Monitoring Program Socioeconomic Monitoring Component: Summary Findings for CNMI, 2016*. U.S. Department of Commerce, NOAA Technical Memorandum, NOAA-TM-NOS-CRCP-34, 69 pp. + Appendices. [PAC]
- Gravinese, P. M. (2018). Ocean acidification impacts the embryonic development and hatching success of the Florida stone crab, *Menippe mercenaria*. *Journal of Experimental Marine Biology and Ecology*, 500, 140–146. <https://doi.org/10.1016/j.jembe.2017.09.001> [FLC]
- Gravinese, P. M., Enochs, I. C., Manzello, D. P., & van Woesik, R. (2018). Warming and  $p\text{CO}_2$  effects on Florida stone crab larvae. *Estuarine, Coastal and Shelf Science*, 204, 193–201. <https://doi.org/10.1016/j.ecss.2018.02.021> [FLC]
- Green, M. A., Waldbusser, G. G., Hubazc, L., Cathcart, E., & Hall, J. (2013). Carbonate mineral saturation state as the recruitment cue for settling bivalves in marine muds. *Estuaries and Coasts*, 36(1), 18–27. [NE]
- Greene, C. M., Blackhart, K., Nohner, J., Candelmo, A., & Nelson, D. M. (2015). A national assessment of stressors to estuarine fish habitats in the contiguous USA. *Estuaries and Coasts*, 38(3), 782–799. [MAB]
- Gruber, N., Hauri, C., Lachkar, Z., Loher, D., Frölicher, T. L., & Plattner, G.-K. (2012). Rapid progression of ocean acidification in the California Current System. *Science*, 337(6091), 220–223. <https://doi.org/10.1126/science.1216773> [WC]
- Gruber, N., Landschützer, P., & Lovenduski, N. S. (2019a). The variable Southern Ocean carbon sink. *Annual Review of Marine Science*, 11(1), 16.1–16.28. <https://doi.org/10.1146/annurev-marine-121916-063407> [INTRO][OO]
- Gruber, N., Clement, D., Carter, B. R., Feely, R. A., van Heuven, S., Hoppema, M., et al. (2019b). The oceanic sink for anthropogenic  $\text{CO}_2$  from 1994 to 2007. *Science*, 363, 1193–1199. <https://doi.org/10.1126/science.aau5153> [OO]
- Guilbert, J., Betts, A. K., Rizzo, D. M., Beckage, B., & Bomblies, A. (2015). Characterization of increased persistence and intensity of precipitation in the northeastern United States. *Geophysical Research Letters*, 42(6), 1888–1893. [NE]
- Guinotte, J. M., Orr, J., Cairns, S., Freiwald, A., Morgan, L., & George, R. (2006). Will human-induced changes in seawater chemistry alter the distribution of deep-sea scleractinian corals? *Frontiers in Ecology and the Environment*, 4(3), 141–146. [PAC]
- Haines, T. A. (1981). Acidic precipitation and its consequences for aquatic ecosystems: a review. *Transactions of the American Fisheries Society*, 110(6), 669–707. [GL]
- Hales, B., Strutton, P. G., Saraceno, M., Letelier, R., Takahashi, T., Feely, R., et al. (2012). Satellite-based prediction of  $p\text{CO}_2$  in coastal waters of the eastern North Pacific. *Progress in Oceanography*, 103, 1–15. <https://doi.org/10.1016/j.pcean.2012.03.001> [OO]
- Hall, L. W. (1987). Acidification effects on larval striped bass, *Morone saxatilis* in Chesapeake Bay tributaries: a review. *Water, Air, and Soil Pollution*, 35(1-2), 87–96. [GL]
- Halle, C. M., & Largier, J. L. (2011). Surface circulation downstream of the Point Arena upwelling center. *Continental Shelf Research*, 31(12), 1260–1272. <https://doi.org/10.1016/j.csr.2011.04.007> [WC]
- Hansen, B. W., Andersen, C. M. B., Hansen, P. J., Nielsen, T. G., Vismann, B., & Tiselius, P. (2019). In situ and experimental evidence for effects of elevated pH on protistan and metazoan grazers. *Journal of Plankton Research*, 41(3), 257–271. <https://doi.org/10.1093/plankt/fbz020> [SAG]
- Harada, N. (2016). Review: Potential catastrophic reduction of sea ice in the western Arctic Ocean: Its impact on biogeochemical cycles and marine ecosystems. *Global and Planetary Change*, 136, 1–17. <https://doi.org/10.1016/j.gloplacha.2015.11.005> [ARC]
- Hare, J. A., Morrison, W. E., Nelson, M. W., Stachura, M. M., Teeters, E. J., Griffis, R. B., et al. (2016). A vulnerability assessment of fish and invertebrates to climate change on the northeast U.S. continental shelf. *PLoS ONE*, 11(2), e0146756. <https://doi.org/10.1371/journal.pone.0146756> [MAB][NE]
- Harvell, C. D., Montecino-Latorre, D., Caldwell, J. M.,

- Burt, J. M., Bosley, K., Keller, A., et al. (2019). Disease epidemic and a marine heat wave are associated with the continental-scale collapse of a pivotal predator *Pycnopodia helianthoides*. *Science Advances*, 5(1), eaau7042. <https://doi.org/10.1126/sciadv.aau7042> [WC]
- Harvey, B. P., Gwynn-Jones, D., & Moore, P. J. (2013). Meta-analysis reveals complex marine biological responses to the interactive effects of ocean acidification and warming. *Ecology and Evolution*, 3(4), 1016–1030. <https://doi.org/10.1002/ece3.516> [AK]
- Harvey, C., Garfield, N., Williams, G., Tolimieri, N., Schroeder, I., Hazen, E., et al. (2018). Ecosystem Status Report of the California Current for 2018: A Summary of Ecosystem Indicators Compiled by the California Current Integrated Ecosystem Assessment Team (CCEIA). U.S. Department of Commerce, NOAA Technical Memorandum NMFS-NWFSC-145. <https://doi.org/10.25923/mvhf-yk36> [WC]
- Hasler, C. T., Jeffrey, J. D., Schneider, E. V. C., Hannan, K. D., Tix, J. A., & Suski, C. D. (2018). Biological consequences of weak acidification caused by elevated carbon dioxide in freshwater ecosystems. *Hydrobiologia*, 806, 1–12. <https://doi.org/10.1007/s10750-017-3332-y> [GL]
- Hattenrath-Lehmann, T. K., Smith, J. L., Wallace, R. B., Merlo, L. R., Koch, F., Mittelsdorf, H., et al. (2015). The effects of elevated CO<sub>2</sub> on the growth and toxicity of field populations and cultures of the saxitoxin-producing dinoflagellate, *Alexandrium fundyense*. *Limnology and Oceanography*, 60(1), 198–214. [MAB]
- Hauri, C., Gruber, N., Vogt, M., Doney, S. C., Feely, R. A., Lachkar, Z., et al. (2013). Spatiotemporal variability and long-term trends of OA in the California Current System. *Biogeosciences*, 10(1), 193–216. doi:10.5194/bg-10-193-2013 [WC]
- Hauri, C., Doney, S. C., Takahashi, T., Erickson, M., Jiang, G., & Ducklow H. W. (2015). Two decades of inorganic carbon dynamics along the West Antarctic Peninsula. *Biogeosciences*, 12, 6761–6779. <https://doi.org/10.5194/bg-12-6761-2015> [OO]
- Hauri, C., Danielson, S., McDonnell, A. M. P., Hopcroft, R. R., Winsor, P., Shipton, P., et al. (2018). From sea ice to seals: a moored marine ecosystem observatory in the Arctic. *Ocean Science*, 14, 1423–1433. <https://doi.org/10.5194/os-14-1423-2018> [ARC]
- Hawkes, L. A., Broderick, A. C., Godfrey, M. H., & Godley, B. J. (2009). Climate change and marine turtles. *Endangered Species Research*, 7, 137–154. <https://doi.org/10.3354/esr00198> [PAC]
- Hermann, A. J., Gibson, G. A., Cheng, W., Ortiz, I., Aydin, K., Wang, M. Y., et al. (2019). Projected biophysical conditions of the Bering Sea to 2100 under multiple emission scenarios. *ICES Journal Marine of Science*, 76, 1280–1304. <https://doi.org/10.1093/icesjms/fsz043> [AK]
- Hickey, B. M. (1998). Coastal oceanography of western North America from the tip of Baja California to Vancouver Island. In A. R. Robinson, K. H. Brink (Eds.), *The Sea, the Global Coastal Ocean* (Vol. 11, pp 345-393). New York, NY: John Wiley & Sons. [WC]
- Himes-Cornell, A., & Kasperski, S. (2015). Assessing climate change vulnerability in Alaska's fishing communities. *Fisheries Research*, 162, 1–11. <https://doi.org/10.1016/j.fishres.2014.09.010> [AK]
- Hjort, J., Karjalainen, O., Aalto, J., Westermann, S., Romanovsky, W.V., Nelson, F.E., et al. (2018). Degrading permafrost puts Arctic infrastructure at risk by mid-century. *Nature Communications*, 9, 5147. <https://doi.org/10.1038/s41467-018-07557-4> [ARC]
- Hoag, H. (2017). News: “Nations agree to ban fishing in Arctic Ocean for at least 16 years.” *Science*, 1 December 2017. <https://doi.org/10.1126/science.aar6437> [ARC]
- Hodgson, E. E., Essington, T. E., & Kaplan, I. C. (2016). Extending vulnerability assessment to include life stages considerations. *PLoS ONE*, 11(7), e0158917. <https://doi.org/10.1371/journal.pone.0158917> [WC]
- Hodgson, E. E., Kaplan, I. C., Marshall, K. N., Leonard, J., Essington, T. E., Busch, D. S., et al. (2018). Consequences of spatially variable ocean acidification in the California Current: Lower pH drives strongest declines in benthic species in southern regions while greatest economic impacts occur in northern regions. *Ecological Modelling*, 383, 106–117. <https://doi.org/10.1016/j.ecolmodel.2018.05.018> [WC]
- Hoegh-Guldberg, O., Mumby, P. J., Hooten, A. J., Steneck, R. S., Greenfield, P., Gomez, E., et al. (2007). Coral reefs under rapid climate change



- and ocean acidification. *Science*, 318(5857), 1737–1742. <https://doi.org/10.1126/science.1152509> [PAC]
- Hoegh-Guldberg, O., Poloczanska, E., Skirving, W., & Dove, S. (2017). Coral reef ecosystems under climate change and ocean acidification. *Frontiers in Ecology and the Environment*, 4, 158. <https://doi.org/10.3389/fmars.2017.00158> [OO][PAC]
- Hogarth, W. T. (2006). Endangered and threatened species: final listing determinations for the elkhorn coral and staghorn coral. *Federal Registry*, 71, 26852–26872. [FLC]
- Hohman, R., Hutto, S., Catton, C., & Koe, F. (2019). Sonoma-Mendocino Bull Kelp Recovery Plan. Unpublished report to Greater Farallones National Marine Sanctuary and the California Department of Fish and Wildlife. San Francisco, CA. 166 pp. [WC]
- Hollowed, A. B., Holsman, K. K., Haynie, A. C., Hermann, A. J., Punt, A. E., Aydin, K., et al. (2020). Integrated modeling to evaluate climate change impacts on coupled social-ecological systems in Alaska. *Frontiers in Marine Science*, 6, 775. <https://doi.org/10.3389/fmars.2019.00775> [AK]
- Holsman, K., Samhuri, J., Cook, G., Hazen, E., Olsen, E., Dillard, M., et al. (2017). An ecosystem-based approach to marine risk assessment. *Ecosystem Health and Sustainability*, 3, e01256. <https://doi.org/10.1002/ehs2.1256> [AK]
- Holsman, K. K., Hazen, E. L., Haynie, A., Gourguet, S., Hollowed, A., Bograd, S. J., et al., (2019). Towards climate resiliency in fisheries management. *ICES Journal of Marine Science*, 76, 1368–1378. <https://doi.org/10.1093/icesjms/fsz031> [AK]
- Hu, X., Nuttall, M. F., Wang, H., Yao, H., Staryk, C. J., McCutcheon, M. R., et al. (2018). Seasonal variability of carbonate chemistry and decadal changes in waters of a marine sanctuary in the Northwestern Gulf of Mexico. *Marine Chemistry*, 205, 16–28. <https://doi.org/10.1016/j.marchem.2018.07.006> [SAG]
- Huang, W.-J., Cai, W.-J., Castelao, R. M., Wang, Y., & Lohrenz, S. E. (2013). Effects of a wind-driven cross-shelf large river plume on biological production and CO<sub>2</sub> uptake on the Gulf of Mexico during spring. *Limnology and Oceanography*, 58(5), 1727–1735. <https://doi.org/10.4319/lo.2013.58.5.1727> [SAG]
- Huang, W.-J., Cai, W.-J., Wang, Y., Lohrenz, S. E., & Murrell, M. C. (2015). The carbon dioxide system on the Mississippi River-dominated continental shelf in the northern Gulf of Mexico: 1. Distribution and air-sea CO<sub>2</sub> flux. *Journal of Geophysical Research—Oceans*, 120(3), 1429–1445. <https://doi.org/10.1002/2014JC010498> [SAG]
- Hudson, K. (2018). Virginia Shellfish Aquaculture Situation and Outlook Report. *VIMS Marine Resource Report No. 2018-9*, Virginia Sea Grant Publication #18-3. Retrieved from [https://www.vims.edu/research/units/centerspartners/map/aquaculture/docs\\_aqua/vims\\_mrr\\_2018-9.pdf](https://www.vims.edu/research/units/centerspartners/map/aquaculture/docs_aqua/vims_mrr_2018-9.pdf). [MAB]
- Hurst, T. P., Fernandez, E. R., Mathis, J. T., Miller, J. A., Stinson, C. S., & Ahgeak, E. F. (2012). Resiliency of juvenile walleye pollock to projected levels of OA. *Aquatic Biology*, 17, 247–259. [AK]
- Hurst, T. P., Fernandez, E. R., & Mathis, J. T. (2013). Effects of ocean acidification on hatch size and larval growth of walleye pollock (*Theragra chalcogramma*). *ICES Journal of Marine Science*, 70(4), 812–822. <https://doi.org/10.1093/icesjms/fst053> [INTRO][AK]
- Hurst, T. P., Laurel, B. J., Mathis, J. T., & Tobosa, L. R. (2016). Effects of elevated CO<sub>2</sub> levels on eggs and larvae of a North Pacific flatfish. *ICES Journal of Marine Science*, 73, 981–990. <https://doi.org/10.1093/icesjms/fsv050> [INTRO][AK]
- Hurst, T. P., Laurel, B. J., Hanneman, E., Haines, S. A., & Ottmar, M. L. (2017). Elevated CO<sub>2</sub> does not exacerbate nutritional stress in larvae of a Pacific flatfish. *Fisheries Oceanography*, 26, 336–349. <https://doi.org/10.1111/fog.12195> [AK]
- Hurst, T. P., Copeman, L. A., Haines, S. A., Meredith, S. D., Daniels, K., & Hubbard, K. M. (2019). Elevated CO<sub>2</sub> alters behavior, growth, and lipid composition of Pacific cod larvae. *Marine Environmental Research*, 145, 52–65. [AK]
- Hutto, S. V. (Ed.) (2016). Climate-Smart Adaptation for North-central California Coastal Habitats. *Report of the Climate-Smart Adaptation Working Group of the Greater Farallones*

National Marine Sanctuary Advisory Council.  
San Francisco, CA. 47 pp. [WC]

ICCA (2015). Alaskan Inuit Food Security Conceptual Framework: How to Assess the Arctic from an Inuit Perspective. Summary and Recommendations Report. Inuit Circumpolar Council-Alaska, Anchorage, AK. Retrieved from <https://iccalaska.org/wp-icc/wp-content/uploads/2016/03/Food-Security-Summary-and-Recommendations-Report.pdf> [ARC]

IJC Great Lakes Water Quality Board Public Engagement Work Group (2018). *Second Binational Great Lakes Basin Poll*. Windsor, ON: International Joint Commission, Great Lakes Regional Office. Retrieved from [https://legacy-files.ijc.org/tinymce/uploaded/WQB/WQB\\_Second\\_Poll\\_Report.pdf](https://legacy-files.ijc.org/tinymce/uploaded/WQB/WQB_Second_Poll_Report.pdf) [GL]

IPCC (2019). IPCC Special Report on the Ocean and Cryosphere in a Changing Climate [Pörtner, H.-O., Roberts, D. C., Masson-Delmotte, V., Zhai, P., Tignor, M., Poloczanska, E., et al. (Eds.)]. In press. [INTRO][NAT]

IPCC (2011). Workshop Report of the Intergovernmental Panel on Climate Change Workshop on Impacts of Ocean Acidification on Marine Biology and Ecosystems [Field, C.B., V. Barros, T.F. Stocker, D. Qin, K.J. Mach, G.-K. Plattner, M.D. Mastrandrea, M. Tignor and K.L. Ebi (Eds.)]. IPCC Working Group II Technical Support Unit, Carnegie Institution, Stanford, California, United States of America, 164 pp. [INTRO][NAT]

Ivanina, A. V., Dickinson, G. H., Matoo, O. B., Bagwe, R., Dickinson, A., Beniash, E., & Sokolova, I. M. (2013). Interactive effects of elevated temperature and CO<sub>2</sub> levels on energy metabolism and biomineralization of marine bivalves *Crassostrea virginica* and *Mercenaria*. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*, 166(1), 101–111. [MAB]

Jacox, M. G., Moore, A. M., Edwards, C. A., & Fiechter, J. (2014). Spatially resolved upwelling in the California Current System and its connections to climate variability. *Geophysical Research Letters*, 41(9), 3189–3196. <https://doi.org/10.1002/2014gl059589> [WC]

Jepson, M., & Colburn, L. L. (2013). Development of social indicators of fishing community vulnerability and resilience in the U.S. southeast and northeast regions. U.S. Department of Commerce, NOAA Technical Memorandum NMFS-F/SPO-129. [NE]

Jiang, L.-Q., Feely, R. A., Carter, B. R., Greeley, D. J., Gledhill, D. K., & Arzayus, K. M. (2015). Climatological distribution of aragonite saturation state in the global oceans. *Global Biogeochemical Cycles*, 29, 1656–1673. <https://doi.org/10.1002/2015GB005198> [OO]

Jiang, L.-Q., Carter, B. R., Feely, R. A., Lauvset, S. K., & Olsen, A. (2019). Surface ocean pH and buffer capacity: Past, present and future. *Scientific Reports*, 9, 18624. <https://doi.org/10.1038/s41598-019-55039-4> [OO]

Johnson, K. S., Plant, J. N., Coletti, L. J., Jannasch, H. W., Sakamoto, C. M., Riser, S. C., et al. (2017). Biogeochemical sensor performance in the SOCCOM profiling float array. *Journal of Geophysical Research: Oceans*, 122(8), 6416–6436. <https://doi.org/10.1002/2017JC012838> [OO]

Jones, J. M., Passow, U., & Fradkin, S. C. (2018). Characterizing the vulnerability of intertidal organisms in Olympic National Park to ocean acidification. *Elementa*, 6(1), 54. <https://doi.org/10.1525/elementa.312> [WC]

Jönsson, B. F., & Salisbury, J. E. (2016). Episodicity in phytoplankton dynamics in a coastal region. *Geophysical Research Letters*, 43, 5821–5828. <https://doi.org/10.1002/2016GL068683> [OO]

Jönsson, B. F., Salisbury, J. E., & Mahadevan, A. (2011). Large variability in continental shelf production of phytoplankton carbon revealed by satellite. *Biogeosciences*, 8(5), 1213–1223. <https://doi.org/10.5194/bg-8-1213-2011> [OO]

Kaplan, I. C., Williams, G. D., Bond, N. A., Hermann, A. J., & Siedlecki, S. A. (2016). Cloudy with a chance of sardines: forecasting sardine distributions using regional climate models. *Fisheries Oceanography*, 25(1), 15–27. <https://doi.org/10.1111/fog.12131> [WC]

Karp, M. A., Peterson, J. O., Lynch, P. D., Griffis, R. B., Adams, C. F., Arnold, W. S., et al. (2019). Accounting for shifting distributions and changing productivity in the development of scientific advice for fishery management. *ICES Journal of Marine Science*, 76, 1305–1315. <https://doi.org/10.1093/icesjms/fsz048> [AK]

Kavanaugh, M. T., Hales, B., Saraceno, M., Spitz, Y. H., White, A. E., & Letelier, R. M. (2014). Hierarchical and dynamic seascapes: A quantitative framework for scaling pelagic biogeochemistry and ecology. *Progress in Oceanography*, 120, 291–304. [OO]

- Kavanaugh, M. T., Oliver, M. J., Chavez, F. P., Letelier, R. M., Muller-Karger, F. E., & Doney, S. C. (2016). Seascales as a new vernacular for pelagic ocean monitoring, management and conservation. *ICES Journal of Marine Science*, 73(7), 1839–1850. [OO]
- Kawaguchi, S., Ishida, A., King, R., Raymond, B., Waller, N., Constable, A., et al. (2013). Risk maps of Antarctic krill under projected Southern OA. *Nature Climate Change*, 3, 843–847. <https://doi.org/10.1038/nclimate1937> [ARC]
- Kennedy, E. V., Perry, C. T., Halloran, P. R., Iglesias-Prieto, R., Schönberg, C. H. L., Wisshak, M., et al. (2013). Avoiding coral reef functional collapse requires local and global action. *Current Biology*, 23(10), 912–918. <https://doi.org/10.1016/j.cub.2013.04.020> [FLC]
- Kennish, M. J., Bricker, S. B., Dennison, W. C., Glibert, P. M., Livingston, R. J., Moore, K. A., et al. (2007). Barnegat Bay–Little Egg Harbor Estuary: Case study of a highly eutrophic coastal bay system. *Ecological Applications*, 17(sp5), S3–S16. <https://doi.org/10.1890/05-0800.1> [MAB]
- Kennish, M. J., Sakowicz, G. P., & Fertig, B. (2016). Recent trends of *Zostera marina* (eelgrass) in a highly eutrophic coastal lagoon in the mid-Atlantic region (USA). *Open Journal of Ecology*, 6(05), 243. [MAB]
- Keppel, E. A., Scrosati, R. A., & Courtenay, S. C. (2012). Ocean acidification decreases growth and development in American lobster (*Homarus americanus*) larvae. *Journal of Northwest Atlantic Fishery Science*, 44, 61–66. [NE]
- Khatiwala, S., Tanhua, T., Mikaloff Fletcher, S., Gerber, M., Doney, S. C., Graven, H. D., et al. (2013). Global ocean storage of anthropogenic carbon. *Biogeosciences*, 10, 2169–2191. <https://doi.org/10.5194/bg-10-2169-2013> [INTRO][OO]
- Kittinger, J. N., Finkbeiner, E. M., Glazier, E. W., & Crowder, L. B. (2012). Human dimensions of coral reef social-ecological systems. *Ecology and Society*, 17(4), 17. <https://doi.org/10.5751/ES-05115-170417> [WC]
- Kleiber, D., Kotowicz, D. M., & Hospital, J. (2018). *Applying National Community Social Vulnerability Indicators to Fishing Communities in the Pacific Island Region*. U.S. Department of Commerce, NOAA Technical Memorandum NMFS-PIFSC-65. <https://doi.org/10.7289/V5/TM-PIFSC-65> [PAC]
- Kleypas, J. A., Castruccio, F. S., Curchitser, E. N., & McLeod, E. (2015). The impact of ENSO on coral heat stress in the western equatorial Pacific. *Global Change Biology*, 21, 2525–2539. <https://doi.org/10.1111/gcb.12881> [OO]
- Kramer, B. J., Davis, T. W., Meyer, K. A., Rosen, B. H., Goleski, J. A., Dick, G. J., et al. (2018). Nitrogen limitation, toxin synthesis potential, and toxicity of cyanobacterial populations in Lake Okeechobee and the St. Lucie River Estuary, Florida, during the 2016 state of emergency event. *PLoS ONE*, 13(5), e0196278. <https://doi.org/10.1371/journal.pone.0196278> [SAG]
- Kroeker, K. J., Kordas, R. L., Crim, R. N., & Singh, G. G. (2010). Meta-analysis reveals negative yet variable effects of ocean acidification on marine organisms. *Ecology Letters*, 13(11), 1419–1434. <https://doi.org/10.1111/j.1461-0248.2010.01518.x> [PAC]
- Kumar, A., AbdElgawad, H., Castellano, I., Lorenti, M., Delledonne, M., Beemster, G. T. S., et al. (2017). Physiological and biochemical analyses shed light on the response of *Sargassum vulgare* to ocean acidification at different timescales. *Frontiers in Plant Science*, 8, 570. <https://doi.org/10.3389/fpls.2017.00570> [FLC]
- Kunz, K. L., Frickenhaus, S., Hardenberg, S., Johansen, T., Leo, E., Pörtner, H.-O., et al. (2016). New encounters in Arctic waters: A comparison of metabolism and performance of polar cod (*Boreogadus saida*) and Atlantic cod (*Gadus morhua*) under ocean acidification and warming. *Polar Biology*, 39(6), 1137–1153. <https://doi.org/10.1007/s00300-016-1932-z> [ARC]
- Lam, V. W. Y., Cheung, W. W. L., & Sumaila, U. R. (2016). Marine capture fisheries in the Arctic: Winners or losers under climate change and ocean acidification? *Fish and Fisheries*, 17, 335–357. <https://doi.org/10.1111/faf.12106> [ARC]
- Landschützer, P., Gruber, N., & Bakker, D. C. E. (2016). Decadal variations and trends of the global ocean carbon sink. *Global Biogeochemical Cycles*, 30(10), 1396–1417. <https://doi.org/10.1002/2015GB005359> [OO]
- Lapointe, G. (2013). Overview of the aquaculture sector in New England. Northeast Regional Ocean Council White Paper. New York, NY: NROC. [NE]
- Laurent, A., Fennel, K., Cai, W.-J., Huang, W.-J., Barbero, L., & Wanninkhof, R. (2017). Eutro-



- phication-induced acidification of coastal waters in the northern Gulf of Mexico: Insights into origin and processes from a coupled physical-biogeochemical model. *Geophysical Research Letters*, 44(2), 946–956. <https://doi.org/10.1002/2016gl071881> [SAG]
- Lauvset, S. K., Key, R. M., Olsen, A., Heuven, S. Van, Velo, A., Lin, X., et al. (2016). A new global interior ocean mapped climatology : the 1° × 1° GLODAP version 2. *Earth System Science Data*, 8, 325–340. <https://doi.org/10.5194/essd-8-325-2016> [PAC]
- Lentz, S. (2003). A climatology of salty intrusions over the continental shelf from Georges Bank to Cape Hatteras. *Journal of Geophysical Research: Oceans*, 108(C10), 3326. [MAB]
- Leong, K. M., Wongbusarakum, S., Ingram, R. J., Mawyer, A., & Poe, M. R. (2019). Improving representation of human well-being and cultural importance in conceptualizing the West Hawai'i ecosystem. *Frontiers in Marine Science*, 6, 231. <https://doi.org/10.3389/fmars.2019.00231> [OO] [PAC]
- Le Quéré, C., Moriarty, R., Andrew, R. M., Canadell, J. G., Sitch, S., Korsbakken, J. I., et al. (2015). Global Carbon Budget 2015. *Earth System Science Data*, 7, 349–396. <https://doi.org/10.5194/essd-7-349-2015>. [INTRO]
- Le Quéré, C., Andrew, R. M., Friedlingstein, P., Sitch, S., Hauck, J., Pongratz, J., et al. (2018). Global Carbon Budget 2018. *Earth System Science Data*, 10, 2141–2194. <https://doi.org/10.5194/essd-10-2141-2018> [INTRO]
- Lessios, H. A. (2016). The great *Diadema antillarum* die-off: 30 years later. *Annual Review of Marine Science*, 8(1), 267–283. <https://doi.org/10.1146/annurev-marine-122414-033857> [FLC]
- Levine, A., Dillard, M., Loerzel, J., & Edwards, P. (2016). *National Coral Reef Monitoring Program Socioeconomic Monitoring Component. Summary Findings for American Samoa, 2014*. U.S. Department of Commerce, NOAA Technical Memorandum CRCP 24, 80 pp. + Appendices. <https://doi.org/10.7289/V5FB50Z1> [PAC]
- Li, H., & Ilyina, T. (2018). Current and future decadal trends in the oceanic carbon uptake are dominated by internal variability. *Geophysical Research Letters*, 45(2), 916–925. <https://doi.org/10.1002/2017GL075370> [WC]
- Li, H., Ilyina, T., Müller, W. A., & Sienz, F. (2016). Decadal predictions of the North Atlantic CO<sub>2</sub> uptake. *Nature Communications*, 7, 11076. <https://doi.org/10.1038/ncomms11076> [OO]
- Li, H., Ilyina, T., Müller, W. A., & Landschützer, P. (2019). Predicting the variable ocean carbon sink. *Science Advances*, 5(4), eaav6471. <https://doi.org/10.1126/sciadv.aav6471> [WC]
- Li, T., Bai, Y., He, X., Xie, Y., Chen, X., Gong, F., & Pan, D. (2018). Satellite-based estimation of particulate organic carbon export in the northern South China Sea. *Journal of Geophysical Research: Oceans*, 123, 8227–8246. <https://doi.org/10.1029/2018JC014201> [OO]
- Lischka, S., Büdenbender, J., Boxhammer, T., & Riebesell, U. (2011). Impact of ocean acidification and elevated temperatures on early juveniles of the polar shelled pteropod *Limacina helicina*: Mortality, shell degradation, and shell growth. *Biogeosciences*, 8, 919–932. <https://doi.org/10.5194/bg-8-919-2011> [ARC]
- Lohrenz, S. E., Cai, W.-J., Chen, F., Chen, X., & Tuel, M. (2010). Seasonal variability in air-sea fluxes of CO<sub>2</sub> in a river-influenced coastal margin. *Journal of Geophysical Research*, 115(C10), C10034. <https://doi.org/10.1029/2009jc005608> [SAG]
- Lohrenz, S. E., Cai, W.-J., Chakraborty, S., Huang, W.-J., Guo, X., He, R., et al. (2018). Satellite estimation of coastal pCO<sub>2</sub> and air-sea flux of carbon dioxide in the northern Gulf of Mexico. *Remote Sensing of Environment*, 207, 71–83. <https://doi.org/10.1016/j.rse.2017.12.039> [SAG]
- Long, W. C., Swiney, K. M., & Foy, R. J. (2013a). Effects of ocean acidification on the embryos and larvae of red king crab, *Paralithodes camtschaticus*. *Marine Pollution Bulletin*, 69, 38–47. <https://doi.org/10.1016/j.marpolbul.2013.01.011> [AK][ARC]
- Long, W. C., Swiney, K. M., Harric, C., Page, H. N., & Foy, R. J. (2013b). Effects of ocean acidification on juvenile red king crab (*Paralithodes camtschaticus*) and Tanner crab (*Chionoecetes bairdi*) growth, condition, calcification, and survival. *PLoS ONE*, 8(4), e360959. <https://doi.org/10.1371/journal.pone.0060959> [AK][ARC]
- Long, W. C., Swiney, K. M., & Foy, R. J. (2016). Effects of high pCO<sub>2</sub> on Tanner crab reproduction and early life history, Part II: carryover effects on larvae from oogenesis and embryogenesis

- are stronger than direct effects. *ICES Journal of Marine Science*, 73(3), 836–848. <https://doi.org/10.1093/icesjms/fsv251> [AK]
- Long, W. C., Van Sant, S. B., Swiney, K. M., & Foy, R. J. (2017). Survival, growth, and morphology of blue king crabs: effect of ocean acidification decreases with exposure time. *ICES Journal of Marine Science*, 74, 1033–1041. <https://doi.org/10.1093/icesjms/fsw197> [AK]
- Lonthair, J., Ern, R., & Esbaugh, A. J. (2017). The early life stages of an estuarine fish, the red drum (*Sciaenops ocellatus*), are tolerant to high  $p\text{CO}_2$ . *ICES Journal of Marine Science*, 74(4), 1042–1050. <https://doi.org/10.1093/icesjms/fsw225> [MAB]
- Lougheed, V. L., Tweedie, C. E., Andresen, C. G., Armendariz, A. M., Escarzaga, S. M., & Tarin, G. (2020). Patterns and drivers of carbon dioxide concentrations in aquatic ecosystems of the Arctic coastal tundra. *Global Biogeochemical Cycles*, 34, e2020GB006552. <https://doi.org/10.1029/2020GB006552> [ARC]
- Lovett, H. B., Snider, S. B., Gore, K. R., & Muñoz, R. C. (Eds.) (2016). Gulf of Mexico Regional Action Plan to Implement the NOAA Fisheries Climate Science Strategy. NOAA Technical Memorandum NMFS-SEFSC-699, 40 pp. [SAG]
- Lowry, L., & Frost, K. (1981). Feeding and trophic relationships of phocid seals and walrus in the eastern Bering Sea. In *The Eastern Bering Sea Shelf: Oceanography and Resources*. (Vol. 2, pp. 813–824). Juneau, AK: NOAA Office of Marine Pollution Assessment. [ARC]
- Lunden, J. J., Georgian, S. E., & Cordes, E. E. (2013). Aragonite saturation states at cold-water coral reefs structured by *Lophelia pertusa* in the northern Gulf of Mexico. *Limnology and Oceanography*, 58, 354–362. <https://doi.org/10.4319/lo.2013.58.1.0354> [SAG]
- Mackie, G. L., & Claudi, R. (2009). *Monitoring and Control of Macrofouling Mollusks in Fresh Water Systems*. Boca Raton, FL: CRC Press. [GL]
- Madge, L., Hospital, J., & Williams, E. T. (2016). *Attitudes and Preferences of Hawaii Non-commercial Fishers: Report from the 2015 Hawaii Saltwater Angler Survey*. U.S. Department of Commerce, NOAA Tech. Memo., NOAA-TM-NMFS-PIFSC-58, 36 pp. + Appendices. <https://doi.org/10.7289/V5/TM-PIFSC-58> [PAC]
- Manizza, M., Follows, M. J., Dutkiewicz, S., Menemenlis, D., McClelland, J. W., Hill, C. N., et al. (2011). A model of the Arctic Ocean carbon cycle. *Journal of Geophysical Research: Oceans*, 116, C12020. <https://doi.org/10.1029/2011JC006998> [ARC]
- Manzello, D., Enochs, I., Musielewicz, S., Carlton, R., & Gledhill, D. (2013). Tropical cyclones cause  $\text{CaCO}_3$  undersaturation of coral reef seawater in a high- $\text{CO}_2$  world. *Journal of Geophysical Research: Oceans*, 118, 5312–5321. <https://doi.org/10.1002/jgrc.20378> [FLC]
- Manzello, D. P. (2010). Ocean acidification hot spots: Spatiotemporal dynamics of the seawater  $\text{CO}_2$  system of eastern Pacific coral reefs. *Limnology and Oceanography*, 55, 239–248. <https://doi.org/10.4319/lo.2010.55.1.0239> [FLC]
- Manzello, D. P., Enochs, I. C., Melo, N., Gledhill, D. K., & Johns, E. M. (2012). Ocean acidification refugia of the Florida Reef Tract. *PLoS ONE*, 7, e41715. <https://doi.org/10.1371/journal.pone.0041715> [FLC]
- Marshall, K. N., Kaplan, I. C., Hodgson, E. E., Hermann, A., Busch, D. S., McElhany, P., et al. (2017). Risks of ocean acidification in the California Current food web and fisheries: ecosystem model projections. *Global Change Biology*, 23(4), 1525–1539. <https://doi.org/10.1111/gcb.13594> [INTRO][ARC][WC][PAC]
- Martin Associates (2018). Economic impacts of maritime shipping in the Great Lakes – St. Lawrence Region. Lancaster, PA. Retrieved from [https://greatlakes-seaway.com/wp-content/uploads/2019/10/eco\\_impact\\_full.pdf](https://greatlakes-seaway.com/wp-content/uploads/2019/10/eco_impact_full.pdf) [GL]
- Mathis, J. T., Hansell, D. A., & Bates, N. R. (2009). Interannual variability of dissolved inorganic carbon distribution and net community production during the Western Arctic Shelf-Basin Interactions Project. *Deep-Sea Research Part II*, 56, 1213–1222. [ARC]
- Mathis, J. T., Byrne, R. H., McNeil, C. L., Pickart, R. P., Juranek, L., Liu, S., et al. (2012). Storm-induced upwelling of high  $p\text{CO}_2$  waters onto the continental shelf of the western Arctic Ocean and implications for carbonate mineral saturation states. *Geophysical Research Letters*, 39, L07606. <https://doi.org/10.1029/2012GL051574> [ARC]
- Mathis, J. T., Cooley, S. R., Lucey, N., Colt, S., Ekstrom, J., Hurst, T., et al. (2015a). Ocean acidification risk assessment for Alaska's fishery

- sector. *Progress in Oceanography*, 136, 71–91. <https://doi.org/10.1016/j.pocean.2014.07.001> [AK] [ARC]
- Matsumoto, K., Tokos, K. S., & Gregory, C. (2015). Ventilation and dissolved oxygen cycle in Lake Superior: Insights from a numerical model. *Geochemistry, Geophysics, Geosystems*, 16(9), 3097–3110. <https://doi.org/10.1002/2015GC005916> [GL]
- McCarthy, K. T., Pichler, T., & Price, R. E. (2005). Geochemistry of Champagne Hot Springs shallow hydrothermal vent field and associated sediments, Dominica, Lesser Antilles. *Chemical Geology*, 224(1-3), 55–68. <https://doi.org/10.1016/j.chemgeo.2005.07.014> [FLC]
- McClatchie, S., Thompson, A. R., Alin, S. R., Siedlecki, S., Watson, W., & Bograd, S. J. (2016). The influence of Pacific Equatorial Water on fish diversity in the southern California Current System. *Journal of Geophysical Research—Oceans*, 121(8), 6121–6136. <https://doi.org/10.1002/2016JC011672> [OO]
- McCutcheon, M. R., Staryk, C. J., & Hu, X. (2019). Characteristics of the carbonate system in a semiarid estuary that experiences summertime hypoxia. *Estuaries and Coasts*, 42, 1509–1523. <https://doi.org/10.1007/s12237-019-00588-0> [SAG]
- McDowell Group (2017). The economic value of Alaska's seafood industry. Anchorage, AK: Alaska Seafood Marketing Institute, 38 pp. Retrieved from <https://uploads.alaskaseafood.org/2017/12/AK-Seafood-Impacts-September-2017.pdf> [AK]
- McElhany, P. (2016). CO<sub>2</sub> sensitivity experiments are not sufficient to show an effect of OA. *ICES Journal of Marine Science*, 74(4), 926–928. Doi:10.1093/icesjms/fsw085 [WC]
- McGuire, A. D., Anderson, L. G., Christensen, T. R., Dallimore, S., Guo, L., Hayes, D.J., et al. (2009). Sensitivity of the carbon cycle in the Arctic to climate change. *Ecological Monographs*, 79(4), 523–555. <https://doi.org/10.1890/08-2025.1> [ARC]
- McLaskey, A. K., Keister, J. E., McElhany, P., Olson, M. B., Busch, D. S., Maher, M., & Winans, A. K. (2016). Development of *Euphausia pacifica* (krill) larvae is impaired under pCO<sub>2</sub> levels currently observed in the northeast Pacific. *Marine Ecology Progress Series*, 555, 65–78. <https://doi.org/10.3354/meps11839> [ARC][WC]
- McManus, M. C., Hare, J. A., Richardson, D. E., & Collie, J. S. (2018). Tracking shifts in Atlantic mackerel (*Scomber scombrus*) larval habitat suitability on the northeast U.S. continental shelf. *Fisheries Oceanography*, 27(1), 49–62. <https://doi.org/10.1111/fog.12233> [MAB]
- Meier, K. J. S., Beaufort, L., Heussner, S., & Ziveri, P. (2014). The role of ocean acidification in *Emiliana huxleyi* coccolith thinning in the Mediterranean Sea. *Biogeosciences*, 11, 2857–2869. <https://doi.org/10.5194/bg-11-2857-2014> [INTRO]
- Meinig, C., Jenkins, R., Lawrence-Slavas, N., & Tabisola, H. (2015). The use of Saldrones to examine spring conditions in the Bering Sea: Vehicle specification and mission performance. *Oceans 2015 MTS/IEEE*, Marine Technology Society and Institute of Electrical and Electronics Engineers, Washington, DC, 19–22 October 2015. [OO]
- Meseck, S. L., Alix, J. H., Swiney, K. M., Long, W. C., Wikfors, G. H., & Foy, R. J. (2016). Ocean acidification affects hemocyte physiology in the Tanner crab (*Chionoecetes bairdi*). *PLoS ONE*, 11(2), e0148477. <https://doi.org/10.1371/journal.pone.0148477> [AK]
- Meseck, S. L., Mercaldo-Allen, R., Kuropat, C., Clark, P., & Goldberg, R. (2018). Variability in sediment-water carbonate chemistry and bivalve abundance after bivalve settlement in Long Island Sound, Milford, Connecticut. *Marine Pollution Bulletin*, 135, 165–175. [NE]
- Metcalf, V. (2015). A Business Plan for Sustainability 2015–2020. Eskimo Walrus Commission, Nome, AK, 19 pp. Retrieved from <https://eskimowalruscommission.org/wp-content/uploads/2019/05/FINAL-EWC-Business-Plan-1-6-16.pdf>. [ARC]
- Meyers, M. T., Cochlan, W. P., Carpenter, E. J., & Kimmerer, W. J. (2019). Effect of ocean acidification on the nutritional quality of marine phytoplankton for copepod reproduction. *PLoS ONE*, 14(5), e0217047. <https://doi.org/10.1371/journal.pone.0217047> [INTRO]
- Michigan Sea Grant College Program (2011). Vital to Our Nation's Economy: Great Lakes Job Report. Michigan Sea Grant, Ann Arbor, MI. Retrieved from <http://www.miseagrant.umich.edu/downloads/economy/11-203-Great-Lakes-Jobs-report.pdf>. [GL]
- Miller, A. W., Reynolds, A. C., Sobrino, C., & Riedel,



- G. F. (2009). Shellfish face uncertain future in high CO<sub>2</sub> world: influence of acidification on oyster larvae calcification and growth in estuaries. *PLoS ONE*, 4(5), e5661. [MAB]
- Miller, J. J., Maher, M., Bohaboy, E., Friedman, C. S., & McElhany, P. (2016). Exposure to low pH reduces survival and delays development in early life stages of Dungeness crab (*Cancer magister*). *Marine Biology*, 163(5), 118. <https://doi.org/10.1007/s00227-016-2883-1> [WC]
- Miller, L. A., Macdonald, R. W., McLaughlin, F., Mucci, A., Yamamoto-Kawai, M., Giesbrecht, K. E., & Williams, W. J. (2014). Changes in the marine carbonate system of the western Arctic: patterns in a rescued data set. *Polar Research*, 33, 20577. <https://doi.org/10.3402/polar.v33.20577> [ARC]
- Miner, C. M., Burnaford, J. L., Ambrose, R. F., Antrim, L., Bohlmann, H., Blanchette, C. A., et al. (2018). Large-scale impacts of sea star wasting disease (SSWD) on intertidal sea stars and implications for recovery. *PLoS ONE*, 13(3), e0192870. <https://doi.org/10.1371/journal.pone.0192870> [WC]
- Moberg, F., & Folke, C. (1999). Ecological goods and services of coral reef ecosystems. *Ecological Economics*, 29, 215–233. [https://doi.org/10.1016/S0921-8009\(99\)00009-9](https://doi.org/10.1016/S0921-8009(99)00009-9) [PAC]
- Mooij, W. M., Hülsmann, S., De Senerpont Domis, L. N., Nolet, B. A., Bodelier, P. L. E., Boers, P. C. M., et al. (2005). The impact of climate change on lakes in the Netherlands: A review. *Aquatic Ecology*, 39(4), 381–400. <https://doi.org/10.1007/s10452-005-9008-0> [GL]
- Moore, S. E., & Grebmeier, J. M. (2018). The Distributed Biological Observatory: Linking physics to biology in the Pacific Arctic region. *Arctic*, 71(5), Suppl. 1. <https://doi.org/10.14430/arctic4606> [ARC]
- Moore, S. E., & Gulland, F. M. D. (2014). Linking marine mammal and ocean health in the 'New Normal' arctic. *Ocean and Coastal Management*, 102(A), 55–57. <https://doi.org/10.1016/j.ocecoaman.2014.08.011> [ARC]
- Muller-Karger, F. E., Smith, J. P., Werner, S., Chen, R., Roffer, M., Liu, Y., et al. (2015). Natural variability of surface oceanographic conditions in the offshore Gulf of Mexico. *Progress in Oceanography*, 134, 54–76. <https://doi.org/10.1016/j.pocean.2014.12.007> [SAG]
- Munroe, D., Narváez, D., Hennen, D., Jacobson, L., Mann, R., Hofmann, E., et al. (2016). Fishing and bottom water temperature as drivers of change in maximum shell length in Atlantic surfclams (*Spisula solidissima*). *Estuarine, Coastal and Shelf Science*, 170, 112–122. [MAB]
- Nagelkerken, I., & Connell, S. D. (2015). Global alteration of ocean ecosystem functioning due to increasing human CO<sub>2</sub> emissions. *Proceedings of the National Academy of Sciences of the United States of America*, 112(43), 13272–13277. <https://doi.org/10.1073/pnas.1510856112> [PAC]
- Nam, S., Kim, H.-J., & Send, U. (2011). Amplification of hypoxic and acidic events by La Niña conditions on the continental shelf off California. *Geophysical Research Letters*, 38, L22602. <https://doi.org/10.1029/2011GL049549> [NAT]
- Narváez, D. A., Munroe, D. M., Hofmann, E. E., Klinck, J. M., Powell, E. N., Mann, R., & Curchitser, E. (2015). Long-term dynamics in Atlantic surfclam (*Spisula solidissima*) populations: The role of bottom water temperature. *Journal of Marine Systems*, 141, 136–148. [MAB]
- NEFSC (2018). State of the Ecosystem—Mid-Atlantic Bight. [https://static1.squarespace.com/static/511cdc7fe4b00307a2628ac6/t/5a9988cc9140b78237c02c82/1520011489334/SOE\\_MAB\\_2018.pdf](https://static1.squarespace.com/static/511cdc7fe4b00307a2628ac6/t/5a9988cc9140b78237c02c82/1520011489334/SOE_MAB_2018.pdf). [MAB]
- NOAA (2014). NOAA's Arctic Action Plan – Supporting the National Strategy for the Arctic Region. U.S. Department of Commerce, National Oceanic and Atmospheric Administration, Silver Spring, MD. 30 pp. Available at: <https://arctic.noaa.gov/Arctic-News/ArtMID/5556/ArticleID/308/NOAAs-Arctic-Action-Plan> [ARC]
- NOAA (2019). NOAA Report on the U.S. Ocean and Great Lakes Economy. Charleston, SC: NOAA Office for Coastal Management. Retrieved from <https://coast.noaa.gov/digitalcoast/training/econreport.html> [GL]
- NOAA Fisheries (2016). Western Regional Action Plan (WRAP), NOAA Fisheries Climate Science Strategy. U.S. Department of Commerce, NOAA Technical Memorandum NMFS-SWFSC-565. 75 pp. <https://doi.org/10.7289/V5/TM-SWFSC-565> [WC]
- NOAA Fisheries (2017). Saltwater Recreational Fisheries in the Mid-Atlantic. Retrieved from

- [ftp://ftp.library.noaa.gov/noaa\\_documents.lib/NMFS/OfcSustainableFisheries/midatlantic-rec-snapshot-2017.pdf](ftp://ftp.library.noaa.gov/noaa_documents.lib/NMFS/OfcSustainableFisheries/midatlantic-rec-snapshot-2017.pdf). [MAB]
- NOAA Fisheries (2019a). Western Regional Implementation Plan (WRIP), NOAA
- Fisheries Ecosystem-Based Fisheries Management Road Map. U.S. Department of Commerce. 25 pp. Retrieved from <https://www.fisheries.noaa.gov/national/ecosystems/ecosystem-based-fishery-management-implementation-plans> [WC]
- NOAA Fisheries (2019b). Landings. Retrieved from <https://foss.nmfs.noaa.gov/apexfoss/f?p=215:200:9482349827593::NO::> [MAB] [NE]
- NOAA OA Steering Committee (2010). NOAA Ocean and Great Lakes Acidification Research Plan. NOAA Special Report, 143 pp. [GL]
- NPFMC (2009). Fishery management plan for fish resources of the Arctic management area. Anchorage, AK: North Pacific Fishery Management Council, 146 pp. [ARC]
- NRC (1992). Global environmental change: Understanding the human dimensions. National Research Council. Washington, DC: National Academies Press <https://doi.org/10.17226/1792> [GL]
- ONMS (2019). Socioeconomics. NOAA/NOS/Office of National Marine Sanctuaries. Retrieved from <https://sanctuaries.noaa.gov/science/socioeconomic/> [WC]
- Olafsson, J., Olafsdottir, S. R., Benoit-Cattin, A., Danielsen, M., Arnarson, T.S., & Takahashi, T. (2009). Rate of Iceland Sea acidification from time series measurements. *Biogeosciences*, 6, 2661–2668. <https://doi.org/10.5194/bg-6-2661-2009> [ARC]
- Olsen, A., Key, R. M., van Heuven, S., Lauvset, S. K., Velo, A., Lin, X., et al. (2016). The global ocean data analysis project version 2 (GLODAPv2)—An internally consistent data product for the world ocean. *Earth System Science Data*, 8(2), 297–323. <https://doi.org/10.5194/essd-8-297-2016> [OO]
- Orensanz, J. L., Ernst, B., Armstrong, D. A., Stabeno P. J., & Livingston, P. (2004). Contraction of the geographic range of distribution of snow crab (*Chionoecetes opilio*) in the eastern Bering Sea: An environmental ratchet? *CalCOFI Rep.*, 45, 65–79. [ARC]
- Orr, J. C., Fabry, V. J., Aumont, O., Bopp, L., Doney, S. C., Feely, R. A., et al. (2005). Anthropogenic ocean acidification over the twenty-first century and its impact on calcifying organisms. *Nature*, 437, 681–686. <https://doi.org/10.1038/nature04095> [INTRO][NAT][OO]
- Osborne, E. B., Thunell, R. C., Marshall, B. J., Holm, J. A., Tappa, E. J., Benitez-Nelson, C., et al. (2016). Calcification of the planktonic foraminifera *Globigerina bulloides* and carbonate ion concentration: Results from the Santa Barbara Basin. *Paleoceanography*, 31(8), 1083–1102. <https://doi.org/10.1002/2016pa002933> [INTRO][WC]
- Osborne, E. B., Thunell, R. C., Gruber, N., Feely, R. A., & Benitez-Nelson, C. R. (2019). Decadal variability in twentieth-century ocean acidification in the California Current Ecosystem. *Nature Geoscience*, 13(1), 43–49. <https://doi.org/10.1038/s41561-019-0499-z>. [INTRO][NAT] [WC]
- Ou, M., Hamilton, T. J., Eom, J., Lyall, E. M., Gallup, J., Jiang, A., et al. (2015). Responses of pink salmon to CO<sub>2</sub>-induced aquatic acidification. *Nature Climate Change*, 5(10), 950–955. <https://doi.org/10.1038/nclimate2694> [AK]
- Pacella, S. R., Brown, C. A., Waldbusser, G. G., Labiosa, R. G., & Hales, B. (2018). Seagrass habitat metabolism increases short-term extremes and long-term offset of CO<sub>2</sub> under future ocean acidification. *Proceedings of the National Academy of Sciences*, 115, 3870–3875. [WC]
- Paerl, H. W., & Huisman, J. (2008). Blooms like it hot. *Science*, 320(5872), 57–58. [GL]
- Park, J. Y., Stock, C. A., Yang, X., Dunne, J. P., Rosati, A., John, J., & Zhang, S. (2018). Modeling global ocean biogeochemistry with physical data assimilation: A pragmatic solution to the equatorial instability. *Journal of Advances in Modeling Earth Systems*, 10(3), 891–906. [OO]
- Parkinson, J. E., Banaszak, A. T., Altman, N. S., LaJeunesse, T. C., & Baums, I. B. (2015). Intraspecific diversity among partners drives functional variation in coral symbioses. *Scientific Reports*, 5, 15667. <https://doi.org/10.1038/srep15667> [FLC]
- Passow, U., & Carlson, C. A. (2012). The biological pump in a high CO<sub>2</sub> world. *Marine Ecology Progress Series*, 470, 249–271. <https://doi.org/10.3354/meps09985> [OO]

- Peeters, F., Livingstone, D. M., Goudsmit, G.-H., Kipfer, R., & Forster, R. (2002). Modeling 50 years of historical temperature profiles in a large central European lake. *Limnology and Oceanography*, 47(1), 186–197. [GL]
- Pendleton, L., Comte, A., Langdon, C., Ekstrom, J. A., Cooley, S. R., Suatoni, L., et al. (2016). Coral reefs and people in a high-CO<sub>2</sub> world: Where can science make a difference to people? *PLoS ONE*, 11(11), e0164699. <https://doi.org/10.1371/journal.pone.0164699> [FLC]
- Perry, C. T., Edinger, E. N., Kench, P. S., Murphy, G. N., Smithers, S. G., Steneck, R. S., & Mumby, P. J. (2012). Estimating rates of biologically driven coral reef framework production and erosion: a new census-based carbonate budget methodology and applications to the reefs of Bonaire. *Coral Reefs*, 31(3), 853–868. <https://doi.org/10.1007/s00338-012-0901-4> [FLC]
- Perry, C. T., Murphy, G. N., Kench, P. S., Smithers, S. G., Edinger, E. N., Steneck, R. S., & Mumby, P. J. (2013). Caribbean-wide decline in carbonate production threatens coral reef growth. *Nature Communications*, 4, 1402. <https://doi.org/10.1038/ncomms2409> [FLC]
- Perry, D. M., Redman, D. H., Widman Jr., J. C., Meseck, S., King, A., & Pereira, J. J. (2015). Effect of ocean acidification on growth and otolith condition of juvenile scup, *Stenotomus chrysops*. *Ecology and Evolution*, 5(18), 4187–4196. <https://doi.org/10.1002/ece3.1678> [MAB]
- Pershing, A. J., Alexander, M. A., Hernandez, C. M., Kerr, L. A., Le Bris, A., Mills, K. E., et al. (2015). Slow adaptation in the face of rapid warming leads to collapse of the Gulf of Maine cod fishery. *Science*, 350(6262), 809–812. <https://doi.org/10.1126/science.aac9819> [NE]
- Peterson, W. T., Fisher, J. L., Strub, P. T., Du, X., Risien, C., Peterson, J., & Shaw, C. T. (2017). The pelagic ecosystem in the Northern California Current off Oregon during the 2014–2016 warm anomalies within the context of the past 20 years. *Journal of Geophysical Research: Oceans*, 122, 7267–7290. <https://doi.org/10.1002/2017JC012952> [OO]
- Phillips, B. F., & Kittaka, J. (2000). *Spiny Lobsters: Fisheries and Culture, Second Edition*. Wiley, 704 pp. <https://doi.org/10.1002/9780470698808> [FLC]
- Phillips, J. C., McKinley, G. A., Bennington, V., Bootsma, H. A., Pilcher, D. J., Sterner, R. W., & Urban, N. R. (2015). The potential for CO<sub>2</sub>-induced acidification in freshwater: A Great Lakes case study. *Oceanography*, 28(2), 136–145. [INTRO][NAT][GL]
- Pilcher, D. J., McKinley, G. A., Kralj, J., Bootsma, H. A., & Reavie, E. D. (2017). Modeled sensitivity of Lake Michigan productivity and zooplankton to changing nutrient concentrations and quagga mussels. *Journal of Geophysical Research: Biogeosciences*, 122(8), 2017–2032. <https://doi.org/10.1002/2017JG003818> [GL]
- Pilcher, D. J., Naiman, D. M., Cross, J. N., Hermann, A. J., Siedlecki, S. A., Gibson, G. A., & Mathis, J. T. (2019). Modeled effect of coastal biogeochemical processes, climate variability, and ocean acidification on aragonite saturation state in the Bering Sea. *Frontiers in Marine Science*, 5, 508. <https://doi.org/10.3389/fmars.2018.00508> [AK] [ARC]
- Polovina, J., Dreflak, K., Baker, J., Bloom, S., Brooke, S., Chan, V., et al. (2016). *Pacific Islands Regional Action Plan*. U.S. Department of Commerce, NOAA Technical Memorandum NMFS-PIFSC-59. <https://doi.org/10.7289/V5/TM-PIFSC-59> [PAC]
- Pörtner, H.O. (2012). Integrating climate-related stressor effects on marine organisms: unifying principles linking molecule to ecosystem-level changes. *Marine Ecology Progress Series*, 470, 273–290. doi:10.3354/meps10123. [INTRO]
- Powell, E. N., Ewing, A. M., & Kuykendall, K. M. (2020). Ocean quahogs (*Arctica islandica*) and Atlantic surfclams (*Spisula solidissima*) on the Mid-Atlantic Bight continental shelf and Georges Bank: The death assemblage as a recorder of climate change and the reorganization of the continental shelf benthos. *Palaeogeography, Palaeoclimatology, Palaeoecology*, 537, 109205. <https://doi.org/10.1016/j.palaeo.2019.05.027> [MAB]
- Price, N. N., Hamilton, S. L., Tootell, J. S., & Smith, J. E. (2011). Species-specific consequences of ocean acidification for the calcareous tropical green algae *Halimeda*. *Marine Ecology Progress Series*, 440, 67–78. <https://doi.org/10.3354/meps09309> [PAC]
- Punt, A. E., Poljak, D., Dalton, M. G., & Foy, R. J. (2014). Evaluating the impact of ocean acidification on fishery yields and profits: The example of red king crab in Bristol Bay. *Ecological Modelling*, 285, 39–53. <https://doi.org/10.1016/j.ecolmod.2014.05.027>



- org/10.1016/j.ecolmodel.2014.04.017 [AK]
- Punt, A. E., Foy, R. J., Dalton, M. G., Long, W. C., & Swiney, K. M. (2016). Effects of long-term exposure to ocean acidification conditions on future southern Tanner crab (*Chionoecetes bairdi*) fisheries management. *ICES Journal of Marine Science*, 73(3), 849–864. <https://doi.org/10.1093/icesjms/fsv205> [ARC]
- Punt, A., Dalton, M., & Foy, R. (in review). Multispecies yield and profit when exploitation rates vary spatially and in the face of OA impacts on mortality: an application for the two North Pacific crab stocks. Manuscript submitted for publication. [AK]
- Putnam, H. M., Barott, K. L., Ainsworth, T. D., & Gates, R. D. (2017). The vulnerability and resilience of reef-building corals. *Current Biology*, 27(11), R528–R540. <https://doi.org/10.1016/j.cub.2017.04.047> [INTRO]
- Qi, D., Chen, L., Chen, B., Gao, Z., Zhong, W., & Feely, R. A. (2017). Increase in acidifying water in the western Arctic Ocean. *Nature Climate Change*, 7, 195–199. <https://doi.org/10.1038/nclimate3228> [ARC]
- Rasmussen, L. L., Gawarkiewicz, G., Owens, W. B., & Lozier, M. S. (2005). Slope water, Gulf Stream, and seasonal influences on southern Mid-Atlantic Bight circulation during the fall-winter transition. *Journal of Geophysical Research: Oceans*, 110(C2), C02009. <https://doi.org/10.1029/2004JC002311> [MAB]
- Raven, J. A., Gobler, C. J., & Hansen, P. J. (2020). Dynamic CO<sub>2</sub> and pH levels in coastal, estuarine, and inland waters: Theoretical and observed effects on harmful algal blooms. *Harmful Algae*, 91, 101594. <https://doi.org/10.1016/j.hal.2019.03.012> [NAT]
- Rawlins, M., Bradley, R., & Diaz, H. (2012). Assessment of regional climate model simulation estimates over the northeast United States. *Journal of Geophysical Research: Atmospheres*, 117(D23), D23112. <https://doi.org/10.1029/2012JD018137> [NE]
- Reimer, J. J., Wang, H., Vargas, R., & Cai, W.-J. (2017). Multidecadal fCO<sub>2</sub> increase along the United States southeast coastal margin. *Journal of Geophysical Research: Oceans*, 122(12), 10061–10072. <https://doi.org/10.1002/2017JC013170> [SAG]
- Reum, J. C. P., Alin, S. R., Feely, R. A., Newton, J., Warner, M., & McElhany, P. (2014). Seasonal carbonate chemistry covariation with temperature, oxygen, and salinity in a fjord estuary: Implications for the design of ocean acidification experiments. *PLoS ONE*, 9(2), e89619. <https://doi.org/10.1371/journal.pone.0089619> [WC]
- Reum, J. C. P., Ferriss, B. E., McDonald, P. S., Farrell, D. M., Harvey, C. J., Klinger, T., & Levin, P. S. (2015). Evaluating community impacts of ocean acidification using qualitative network models. *Marine Ecology Progress Series*, 536, 11–24. <https://doi.org/10.3354/meps11417> [ARC]
- Reum, J. C. P., Alin, S. R., Harvey, C. J., Bednaršek, N., Evans, W., Feely, R. A., et al. (2016). Interpretation and design of ocean acidification experiments in upwelling systems in the context of carbonate chemistry co-variation with temperature and oxygen. *ICES Journal of Marine Science*, 73(3), 582–595. <https://doi.org/10.1093/icesjms/fsu231> [WC]
- Reum, J. C. P., Blanchard, J. L., Holsman, K. K., Aydin, K., & Punt, A. E. (2019). Species-specific ontogenetic diet shifts attenuate trophic cascades and lengthen food chains in exploited ecosystems. *Oikos*, 128, 1051–1064. <https://doi.org/10.1111/oik.05630> [AK]
- Reum, J. C. P., Blanchard, J. L., Holsman, K. K., Aydin, K., Hollowed, A. B., Hermann, A. J., et al. (2020). Ensemble projections of future climate change impacts on the eastern Bering Sea food web using a multispecies size spectrum model. *Frontiers in Marine Science*, 7, 124. <https://doi.org/10.3389/fmars.2020.00124> [AK]
- Reyes-Nivia, C., Diaz-Pulido, G., Kline, D., Hoegh-Guldberg, O., & Dove, S. (2013). Ocean acidification and warming scenarios increase microbioerosion of coral skeletons. *Global Change Biology*, 19, 1919–1929. <https://doi.org/10.1111/gcb.12158> [FLC]
- Rheuban, J. E., Doney, S. C., Cooley, S. R., & Hart, D. R. (2018). Projected impacts of future climate change, ocean acidification, and management on the US Atlantic sea scallop (*Placopecten magellanicus*) fishery. *PLoS ONE*, 13(9), e0203536. <https://doi.org/10.1371/journal.pone.0203536> [NE]
- Riebesell, U., Bach, L. T., Bellerby, R. G. J., Bermúdez Monsalve, J. R., Boxhammer, T., Czerny, J., et al. (2017). Competitive fitness of a predominant pelagic calcifier impaired by ocean acidification. *PLoS ONE*, 12(12), e0187441. <https://doi.org/10.1371/journal.pone.0187441> [WC]

- fication. *Nature Geoscience*, 10, 19–23. <https://doi.org/10.1038/ngeo2854> [INTRO]
- Riekenberg, J., Bargu, S., & Twilley, R. (2015). Phytoplankton community shifts and harmful algae presence in a diversion influenced estuary. *Estuaries and Coasts*, 38(6), 2213–2226. <https://doi.org/10.1007/s12237-014-9925-z> [SAG]
- Ries, J. B., Cohen, A. L., & McCorkle, D. C. (2009). Marine calcifiers exhibit mixed responses to CO<sub>2</sub>-induced ocean acidification. *Geology*, 37(12), 1131–1134. <https://doi.org/10.1130/G30210A.1> [NE]
- Ries, J. B., Ghazaleh, M. N., Connolly, B., Westfield, I., & Castillo, K. D. (2016). Impacts of seawater saturation state ( $\Omega_A = 0.4$ –4.6) and temperature (10, 25° C) on the dissolution kinetics of whole-shell biogenic carbonates. *Geochimica et Cosmochimica Acta*, 192, 318–337. <https://doi.org/10.1016/j.gca.2016.07.001> [MAB]
- Riser, S. C., Swift, D., & Drucker, R. (2018). Profiling floats in SOCCOM: Technical capabilities for studying the Southern Ocean. *Journal of Geophysical Research: Oceans*, 123(6), 4055–4073. <https://doi.org/10.1002/2017JC013419>. [WC]
- Rivest, E. B., Comeau, S., & Cornwall, C. E. (2017). The role of natural variability in shaping the response of coral reef organisms to climate change. *Current Climate Change Reports*, 3, 271–281. <https://doi.org/10.1007/s40641-017-0082-x> [FLC]
- Robbins, L. L., Daly, K. L., Barbero, L., Wanninkhof, R., He, R., Zong, H., et al. (2018). Spatial and temporal variability of pCO<sub>2</sub>, carbon fluxes, and saturation state on the West Florida Shelf. *Journal of Geophysical Research: Oceans*, 123, 6174–6188. <https://doi.org/10.1029/2018jc014195> [SAG]
- Roman, M. R., Pierson, J. J., Kimmel, D. G., Boicourt, W. C., & Zhang, X. (2012). Impacts of hypoxia on zooplankton spatial distributions in the northern Gulf of Mexico. *Estuaries and Coasts*, 35(5), 1261–1269 [SAG]
- Ross, E., & Behringer, D. (2019). Changes in temperature, pH, and salinity affect the sheltering responses of Caribbean spiny lobsters to chemosensory cues. *Scientific Reports*, 9(1), 4375. <https://doi.org/10.1038/s41598-019-40832-y> [FLC]
- Rowe, M. D., Anderson, E. J., Vanderploeg, H. A., Pothoven, S. A., Elgin, A. K., Wang, J., & Yousef, F. (2017). Influence of invasive quagga mussels, phosphorus loads, and climate on spatial and temporal patterns of productivity in Lake Michigan: A biophysical modeling study. *Limnology and Oceanography*, 62(6), 2629–2649. <https://doi.org/10.1002/lno.10595> [GL]
- Rykaczewski, R. R., & Dunne, J. P. (2010). Enhanced nutrient supply to the California Current Ecosystem with global warming and increased stratification in an earth system model. *Geophysical Research Letters*, 37(21), L21606. <https://doi.org/10.1029/2010gl045019> [WC]
- Rykaczewski, R. R., Dunne, J. P., Sydeman, W. J., García-Reyes, M., Black, B. A., & Bograd, S. J. (2015). Poleward displacement of coastal upwelling-favorable winds in the ocean's eastern boundary currents through the 21st century. *Geophysical Research Letters*, 42(15), 6424–6431. <https://doi.org/10.1002/2015gl064694> [WC]
- Saba, G. K., Goldsmith, K. A., Cooley, S. R., Grosse, D., Meseck, S. L., Miller, A. W., et al. (2019). Recommended priorities for research on ecological impacts of ocean and coastal acidification in the U.S. Mid-Atlantic. *Estuarine, Coastal and Shelf Science*, 225, 106188. <https://doi.org/10.1016/j.ecss.2019.04.022> [MAB]
- Saba, V. S., Friedrichs, M. A. M., Antoine, D., Armstrong, R. A., Asanuma, I., Behrenfeld, M. J., et al. (2011). An evaluation of ocean color model estimates of marine primary productivity in coastal and pelagic regions across the globe. *Biogeosciences*, 8, 489–503. <https://doi.org/10.5194/bg-8-489-2011> [OO]
- Saba, V. S., Griffies, S. M., Anderson, W. G., Winton, M., Alexander, M. A., Delworth, T. L., et al. (2016). Enhanced warming of the northwest Atlantic Ocean under climate change. *Journal of Geophysical Research: Oceans*, 121(1), 118–132. <https://doi.org/10.1002/2015JC011346> [MAB]
- Sabine, C. L., Feely, R. A., Gruber, N., Key, R. M., Lee, K., Bullister, J. L., et al. (2004). The oceanic sink for anthropogenic CO<sub>2</sub>. *Science*, 305, 367–371. <https://doi.org/10.1126/science.1097403> [OO] [INTRO]
- Sabine, C., Sutton, A., McCabe, K., Lawrence-Slavas, N., Alin, S., Feely, R., et al. (2020). Evaluation of a new autonomous surface vehicle carbon dioxide system. *Journal of Atmospheric and Oceanic Technology*. <https://doi.org/10.1175/>

- Salisbury, J., Green, M., Hunt, C., & Campbell, J. (2008). Coastal acidification by rivers: A threat to shellfish? *Eos, Transactions American Geophysical Union*, 89(50), 513. [NE]
- Salisbury, J., Vandemark, D., Jönsson, B., Balch, W., Chakraborty, S., Lohrenz, S., et al. (2015). How can present and future satellite missions support scientific studies that address ocean acidification? *Oceanography*, 28(2), 108–121. <https://doi.org/10.5670/oceanog.2015.35> [OO] [NAT]
- Salisbury, J. E., & Jönsson, B. F. (2018). Rapid warming and salinity changes in the Gulf of Maine alter surface ocean carbonate parameters and hide ocean acidification. *Biogeochemistry*, 141(3), 401–418. <https://doi.org/10.1007/s10533-018-0505-3> [NE]
- Sanford, E., Sones, J. L., García-Reyes, M., Goddard, J. H. R., & Largier, J. L. (2019). Widespread shifts in the coastal biota of northern California during the 2014–2016 marine heatwaves. *Scientific Reports*, 9, 4216. <https://doi.org/10.1038/s41598-019-40784-3> [OO]
- Schmidt, M., Gerlach, G., Leo, E., Kunz, K. L., Swoboda, S., Pörtner, H. O., et al. (2017). Impact of ocean warming and acidification on the behaviour of two co-occurring gadid species, *Boreogadus saida* and *Gadus morhua*, from Svalbard. *Marine Ecology Progress Series*, 571, 183–191. [ARC]
- Schönberg, C. H. L., Fang, J. K. H., Carreiro-Silva, M., Tribollet, A., & Wisshak, M. (2017). Bioerosion: the other ocean acidification problem. *ICES Journal of Marine Science*, 74, 895–925. <https://doi.org/10.1093/icesjms/fsw254> [FLC]
- Schweitzer, C. C., & Stevens, B. G. (2019). The relationship between fish abundance and benthic community structure on artificial reefs in the Mid-Atlantic Bight, and the importance of sea whip corals *Leptogorgia virgulata*. *PeerJ*, 7, e7277. <https://doi.org/10.7717/peerj.7277> [MAB]
- Semiletov, I. P., Pipko, I. I., Repina, I., & Shakhova, N. E. (2007). Carbonate chemistry dynamics and carbon dioxide fluxes across the atmosphere–ice–water interfaces in the Arctic Ocean: Pacific sector of the Arctic. *Journal of Marine Systems*, 66(1–4), 204–226. <https://doi.org/10.1016/j.jmarsys.2006.05.012> [ARC]
- Seung, C. K., Dalton, M. G., Punt, A. E., Poljak, D., & Foy, R. (2015). Economic impacts of changes in an Alaska crab fishery from OA. *Climate Change Economics*, 6(4), 1550017. <https://doi.org/10.1142/S2010007815500177> [AK] [ARC]
- Shadwick, E. H., Thomas, H., Gratton, Y., Leong, D., Moore, S. A., Papkyriakou, T., & Prowe, A. E. F. (2011). Export of Pacific carbon through the Arctic archipelago to the North Atlantic. *Continental Shelf Research*, 31(7-8), 806–816. <https://doi.org/10.1016/j.csr.2011.01.014> [ARC]
- Shamberger, K. E. F., Cohen, A. L., Golbuu, Y., McCorkle, D. C., Lentz, S. J., & Barkley, H. C. (2014). Diverse coral communities in naturally acidified waters of a Western Pacific reef. *Geophysical Research Letters*, 41(2), 499–504. <https://doi.org/10.1002/2013GL058489> [FLC]
- Shaw, E. C., McNeil, B. I., Tilbrook, B., Matear, R., & Bates, M. L. (2013). Anthropogenic changes to seawater buffer capacity combined with natural reef metabolism induce extreme future coral reef CO<sub>2</sub> conditions. *Global Change Biology*, 19(5), 1632–1641. <https://doi.org/10.1111/gcb.12154> [FLC]
- Sherman, K., Grosslein, M., Mountain, D., Busch, D., O'Reilly, J., & Theroux, R. (1996). The Northeast Shelf ecosystem: An initial perspective. In K. Sherman, N. A. Jaworski, & T. J. Smayda (Eds.), *The Northeast Shelf Ecosystem: Assessment, Sustainability, and Management* (pp. 103–126). Cambridge, MA: Blackwell Science. [MAB][NE]
- Sherr, E. B., & Sherr, B. F. (2002). Significance of predation by protists in aquatic microbial food webs. *Antonie van Leeuwenhoek*, 81, 293–308. <https://doi.org/10.1023/A:1020591307260> [SAG]
- Shi, X., Li, S., Wei, L., Qin, B., & Brookes, J. D. (2017). CO<sub>2</sub> alters community composition of freshwater phytoplankton: A microcosm experiment. *Science of The Total Environment*, 607, 69–77. <https://doi.org/10.1016/j.scitotenv.2017.06.224> [GL]
- Shutler, J. D., Wanninkhof, R., Nightingale, P. D., Woolf, D. K., Bakker, D. C. E., Watson, A., et al. (2020). Satellites will address critical science priorities for quantifying ocean carbon. *Frontiers in Ecology and the Environment*, 18(1), 27–35. <https://doi.org/10.1002/fee.2129> [OO] [NAT]
- Siedlecki, S. A., Kaplan, I. C., Hermann, A. J., Nguy-



- en, T. T., Bond, N. A., Newton, J. A., et al. (2016). Experiments with seasonal forecasts of ocean conditions for the northern region of the California Current upwelling system. *Scientific Reports*, 6, 27203. <https://doi.org/10.1038/srep27203> and <https://www.nature.com/articles/srep27203#supplementary-information> [WC]
- Siedlecki, S. A., Pilcher, D. J., Hermann, A. J., Coyle, K., & Mathis, J. T. (2017). The importance of freshwater to spatial variability of aragonite saturation state in the Gulf of Alaska. *Journal of Geophysical Research: Oceans*, 122, 8482–8502. <https://doi.org/10.1002/2017JC012791> [ARC]
- Siegel, D. A., Buesseler, K. O., Doney, S. C., Sailley, S. F., Behrenfeld, M. J., & Boyd, P. W. (2014). Global assessment of ocean carbon export by combining satellite observations and food-web models. *Global Biogeochemical Cycles*, 28, 181–196. <https://doi.org/10.1002/2013gb004743> [OO]
- Sinha, E., Michalak, A. M., & Balaji, V. (2017). Eutrophication will increase during the 21st century as a result of precipitation changes. *Science*, 357(6349), 405–408. <https://doi.org/10.1126/science.aan2409> [NE]
- Sloyan, B. M., Wilkin, J., Hill, K. L., Chidichimo, M. P., Cronin, M. F., Johannessen, J. A., et al. (2019a). Evolving the physical global ocean observing system for research and application services through international coordination. *Frontiers in Marine Science*, 6, 449. <https://doi.org/10.3389/fmars.2019.00449> [WC]
- Sloyan, B. M., Wanninkhof, R., Kramp, M., Johnson, G. C., Talley, L. D., Tanhua, T., et al. (2019b). The Global Ocean Ship-Based Hydrographic Investigations Program (GO-SHIP): A platform for integrated multidisciplinary ocean science. *Frontiers in Marine Science*, 6, 445. <https://doi.org/10.3389/fmars.2019.00445> [WC]
- Smith, J. E., Brainard, R., Carter, A., Dugas, S., Edwards, C., Harris, J., et al. (2016). Re-evaluating the health of coral reef communities: Baselines and evidence for human impacts across the central Pacific. *Proceedings of the Royal Society B: Biological Sciences*, 283(1822), 20151985. <https://doi.org/10.1098/rspb.2015.1985> [PAC]
- Speights, C. J., Silliman, B. R., & McCoy, M. W. (2017). The effects of elevated temperature and dissolved pCO<sub>2</sub> on a marine foundation species. *Ecology and Evolution*, 7(11), 3808–3814. <https://doi.org/10.1002/ece3.2969> [MAB]
- Steffen, M. M., Davis, T. W., McKay, R. M. L., Bullerjahn, G. S., Krausfeldt, L. E., Stough, J. M. A., et al. (2017). Ecophysiological examination of the Lake Erie Microcystis bloom in 2014: linkages between biology and the water supply shutdown of Toledo, OH. *Environmental Science and Technology*, 51(12), 6745–6755. <https://doi.org/10.1021/acs.est.7b00856> [GL]
- Steinacher, M., Joos, F., Frölicher, T. L., Plattner, G.-K., & Doney, S. C. (2009). Imminent ocean acidification in the Arctic projected with the NCAR global coupled carbon cycle-climate model. *Biogeosciences*, 6, 515–533. <https://doi.org/10.5194/bg-6-515-2009> [ARC][OO]
- Steinberg, D. K., & Landry, M. R. (2017). Zooplankton and the ocean carbon cycle. *Annual Review of Marine Science*, 9(1), 413–444. <https://doi.org/10.1146/annurev-marine-010814-015924> [SAG]
- Steiner, N. S., Christian, J. R., Six, K. D., Yamamoto, A., & Yamamoto-Kawai, M. (2014). Future ocean acidification in the Canada Basin and surrounding Arctic ocean from CMIP5 earth system models. *Journal of Geophysical Research—Oceans*, 119, 332–347. <https://doi.org/10.1002/2013JC009069> [ARC]
- Steiner, N., Deal, C., Lannuzel, D., Lavoie, D., Massonnet, F., Miller, L. A., et al. (2016). What sea-ice biogeochemical modellers need from observers. *Elementa: Science of the Anthropocene*, 4, 00084. <https://doi.org/10.12952/journal.elementa.000084> [ARC]
- Stepien, C.A., M.R. Snyder, & A.E. Elz. (2019). Invasion genetics of the silver carp *Hypophthalmichthys molitrix* across North America: Differentiation of fronts, introgression, and eDNA metabarcoding detection. *PLoS ONE*, 14(3), e0203012. <https://doi.org/10.1371/journal.pone.0203012> [OO]
- Storlazzi, C. D., Reguero, B. G., Cole, A. D., Lowe, E., Shope, J. B., Gibbs, A. E., et al. (2019). Rigorously valuing the role of U.S. coral reefs in coastal hazard risk reduction: U.S. Geological Survey Open-File Report 2019–1027, 42 pp., <https://doi.org/10.3133/ofr20191027>. [PAC][FLC][NAT]
- Stubler, A. D., Furman, B. T., & Peterson, B. J. (2015). Sponge erosion under acidification and

- warming scenarios: differential impacts on living and dead coral. *Global Change Biology*, 21(11), 4006–4020. <https://doi.org/10.1111/gcb.13002> [FLC]
- Sunda, W. G., & Cai, W.-J. (2012). Eutrophication induced CO<sub>2</sub>-acidification of subsurface coastal waters: interactive effects of temperature, salinity, and atmospheric pCO<sub>2</sub>. *Environmental Science and Technology*, 46(19), 10651–10659. <https://doi.org/10.1021/es300626f> [NAT]
- Sutton, A. J., Sabine, C. L., Maenner-Jones, S., Lawrence-Slavas, N., Meinig, C., Feely, R. A., et al. (2014a). A high-frequency atmospheric and seawater pCO<sub>2</sub> data set from 14 open-ocean sites using a moored autonomous system. *Earth System Science Data*, 6(2). <https://doi.org/10.5194/essd-6-353-20142014>. [WC]
- Sutton, A. J., Feely, R. A., Sabine, C. L., McPhaden, M. J., Takahashi, T., Chavez, F. P., et al. (2014b). Natural variability and anthropogenic change in equatorial Pacific surface ocean pCO<sub>2</sub> and pH. *Global Biogeochemical Cycles*, 28, 131–145. <https://doi.org/10.1002/2013GB004679> [PAC]
- Sutton, A. J., Wanninkhof, R., Sabine, C. L., Feely, R. A., Cronin, M. F., & Weller, R. A. (2017). Variability and trends in surface seawater pCO<sub>2</sub> and CO<sub>2</sub> flux in the Pacific Ocean. *Geophysical Research Letters*, 44, 5627–5636. <https://doi.org/10.1002/2017GL073814> [OO]
- Sutton, A. J., Feely, R. A., Maenner-Jones, S., Musielwicz, S., Osborne, J., Dietrich, C., et al. (2019). Autonomous seawater pCO<sub>2</sub> and pH time series from 40 surface buoys and the emergence of anthropogenic trends. *Earth System Science Data*, 11, 421–439. <https://doi.org/10.5194/essd-11-421-2019> [OO][PAC][SAG]
- Swiney, K. M., Long, W. C., & Foy, R. J. (2016). Effects of high pCO<sub>2</sub> on Tanner crab reproduction and early life history—Part I: long-term exposure reduces hatching success and female calcification, and alters embryonic development. *ICES Journal of Marine Science*, 73(3), 825–835. <https://doi.org/10.1093/icesjms/fsv201> [AK][ARC]
- Swiney, K. M., Long, W. C., & Foy, R. J. (2017). Decreased pH and increased temperatures affect young-of-the-year red king crab (*Paralithodes camtschaticus*). *ICES Journal of Marine Science*, 74(4), 1191–1200. <https://doi.org/10.1093/icesjms/fsw251> [AK]
- Sydeman, W. J., García-Reyes, M., Schoeman, D. S., Rykaczewski, R. R., Thompson, S. A., Black, B. A., & Bograd, S. J. (2014). Climate change and wind intensification in coastal upwelling ecosystems. *Science*, 345(6192), 77–80. <https://doi.org/10.1126/science.1251635> [WC]
- Takahashi, T., Sutherland, S. C., Chipman, D. W., Goddard, J. G., Ho, C., Newberger, T., et al. (2014). Climatological distributions of pH, pCO<sub>2</sub>, total CO<sub>2</sub>, alkalinity, and CaCO<sub>3</sub> saturation in the global surface ocean, and temporal changes at selected locations. *Marine Chemistry*, 164, 95–125. [OO]
- Talmage, S. C., & Gobler, C. J. (2009). The effects of elevated carbon dioxide concentrations on the metamorphosis, size, and survival of larval hard clams (*Mercenaria mercenaria*), bay scallops (*Argopecten irradians*), and Eastern oysters (*Crassostrea virginica*). *Limnology and Oceanography*, 54(6), 2072–2080. [MAB]
- Tanhua, T., Jones, E. P., Jeansson, E., Jutterström, S., Smethie Jr., W. M., Wallace, D. W. R., & Anderson, L. G. (2009). Ventilation of the Arctic Ocean: Mean ages and inventories of anthropogenic CO<sub>2</sub> and CFC-11. *Journal of Geophysical Research*, 114, C01002. <https://doi.org/10.1029/2008JC004868.9> [ARC]
- Tanhua, T., Pouliquen, S., Hausman, J., O'Brien, K., Bricher, P., & de Bruin, T., et al. (2019). Ocean fair data services. *Frontiers in Marine Science*, 6, 440. <https://doi.org/10.3389/fmars.2019.00440> [NAT]
- Taylor, K. E., Stouffer, R. J., & Meehl, G. A. (2012). An overview of CMIP5 and the experiment design. *Bulletin of the American Meteorological Society*, 93(4), 485–498. [OO]
- Thibodeau, P. S., Steinberg, D. K., Stammerjohn, S. E., & Hauri, C. (2019). Environmental controls on pteropod biogeography along the Western Antarctic Peninsula. *Limnology and Oceanography*, 64, S240–S256. <https://doi.org/10.1002/lno.11041> [INTRO]
- Thor, P., & Dupont, S. (2015). Transgenerational effects alleviate severe fecundity loss during ocean acidification in a ubiquitous planktonic copepod. *Global Change Biology*, 21(6), 2261–2271. <https://doi.org/10.1111/gcb.12815> [ARC]
- Thor, P., & Oliva, E. O. (2015). Ocean acidification elicits different energetic responses in an Arctic and a boreal population of the copepod *Pseudocalanus acuspes*. *Marine Biology*, 162,

799–807. <https://doi.org/10.1007/s00227-015-2625-9> [ARC]

- Tilbrook, B., Jewett, E. B., DeGrandpre, M. D., Hernandez-Ayon, J. M., Feely, R. A., Gledhill, D. K., et al. (2019). Towards an enhanced ocean acidification observing network: From people to technology to data synthesis and information exchange. *Frontiers in Marine Science*, 6, 337, Oceanobs19: An Ocean of Opportunity. <https://doi.org/10.3389/fmars.2019.00337> [OO]
- Tjiputra, J. F., Olsen, A., Bopp, L., Lenton, A., Pfeil, B., Roy, T., et al. (2014). Long-term surface  $p\text{CO}_2$  trends from observations and models. *Tellus B: Chemical and Physical Meteorology*, 66(1), 23083. [NE]
- Towle, E. K., Enochs, I. C., & Langdon, C. (2015). Threatened Caribbean coral is able to mitigate the adverse effects of ocean acidification on calcification by increasing feeding rate. *PLoS ONE*, 10(4), e0123394. <https://doi.org/10.1371/journal.pone.0123394> [INTRO]
- Townsend, D. W., Thomas, A. C., Mayer, L. M., Thomas, M. A., & Quinlan, J. A. (2006). Oceanography of the northwest Atlantic continental shelf. *The Sea: The Global Coastal Ocean: Interdisciplinary Regional Studies and Syntheses, Vol. 14A*, pp. 119–168. Harvard University Press, A. R. Robinson and K. Brink, Eds. [NE]
- Tribollet, A., Godinot, C., Atkinson, M., & Langdon, C. (2009). Effects of elevated  $p\text{CO}_2$  on dissolution of coral carbonates by microbial euendoliths. *Global Biogeochemical Cycles*, 23(3), GB3008. <https://doi.org/10.1029/2008GB003286> [FLC]
- Trigg, S. A., McElhany, P., Maher, M., Perez, D., Busch, D. S., & Nichols, K. M. (2019). Uncovering mechanisms of global ocean change effects on the Dungeness crab (*Cancer magister*) through metabolomics analysis. *Scientific Reports*, 9(1), 10717. <https://doi.org/10.1038/s41598-019-46947-6> [WC]
- Trolle, D., Hamilton, D. P., Pilditch, C. A., Duggan, I. C., & Jeppesen, E. (2011). Predicting the effects of climate change on trophic status of three morphologically varying lakes: Implications for lake restoration and management. *Environmental Modelling & Software*, 26(4), 354–370. [GL]
- Turi, G., Lachkar, Z., Gruber, N., & Munnich, M. (2016). Climatic modulation of recent trends in ocean acidification in the California Current System. *Environmental Research Letters*, 11, 014007. <https://doi.org/10.1088/1748-9326/11/1/014007> [WC]
- Turk, D., Wang, H., Hu, X., Gledhill, D. K., Wang, Z. A., Jiang, L., & Cai, W.-J. (2019). Time of emergence of surface ocean carbon dioxide trends in the North American coastal margins in support of ocean acidification observing system design. *Frontiers in Marine Science*, 6, 91. <https://doi.org/10.3389/fmars.2019.00091> [OO]
- Uthicke, S., Soars, N., Foo, S., & Byrne, M. (2013). Effects of elevated  $p\text{CO}_2$  and the effect of parent acclimation on development in the tropical Pacific sea urchin *Echinometra mathaei*. *Marine Biology*, 160(8), 1913–1926. <https://doi.org/10.1007/s00227-012-2023-5> [FLC]
- Van Dam, B. R., & Wang, H. (2019). Decadal-scale acidification trends in adjacent North Carolina estuaries: Competing role of anthropogenic  $\text{CO}_2$  and riverine alkalinity loads. *Frontiers in Marine Science*, 6, 136. <https://doi.org/10.3389/fmars.2019.00136> [SAG]
- Van de Waal, D. B., Verspagen, J. M., Finke, J. F., Vournazou, V., Immers, A. K., Kardinaal, W. E. A., et al. (2011). Reversal in competitive dominance of a toxic versus non-toxic cyanobacterium in response to rising  $\text{CO}_2$ . *The ISME Journal*, 5(9), 1438. [GL]
- van Oppen, M. J. H., Oliver, J. K., Putnam, H. M., & Gates, R. D. (2015). Building coral reef resilience through assisted evolution. *Proceedings of the National Academy of Sciences*, 112, 2307–2313. <https://doi.org/10.1073/pnas.1422301112> [FLC]
- van Tussenbroek, B. I., Hernández Arana, H. A., Rodríguez-Martínez, R. E., Espinoza-Avalos, J., Canizales-Flores, H. M., González-Godoy, C. E., et al. (2017). Severe impacts of brown tides caused by *Sargassum* spp. on near-shore Caribbean seagrass communities. *Marine Pollution Bulletin*, 122(1-2), 272–281. <https://doi.org/10.1016/j.marpolbul.2017.06.057> [FLC]
- Vargas, C. A., de la Hoz, M., Aguilera, V., San Martín, V., Manríquez, P. H., Navarro, J. M., et al. (2013).  $\text{CO}_2$ -driven ocean acidification reduces larval feeding efficiency and changes food selectivity in the mollusk *Concholepas concholepas*. *Journal of Plankton Research*, 35, 1059–1068. <https://doi.org/10.1093/plankt/fbt045> [MAB]
- Vargas-Ángel, B., Richards, C. L., Vroom, P. S., Price,



- N. N., Schils, T., Young, C. W., et al. (2015). Baseline assessment of net calcium carbonate accretion rates on U.S. Pacific reefs. *PLoS ONE*, 10(12), e0142196. <https://doi.org/10.1371/journal.pone.0142196> [PAC][FLC]
- Waldbusser, G. G., Powell, E. N., & Mann, R. (2013). Ecosystem effects of shell aggregations and cycling in coastal waters: an example of Chesapeake Bay oyster reefs. *Ecology*, 94(4), 895–903. <https://doi.org/10.1890/12-1179.1> [MAB]
- Waldbusser, G. G., Hales, B., Langdon, C. J., Haley, B. A., Schrader, P., Brunner, E. L., Gray, M. W., et al. (2015). Ocean acidification has multiple modes of action on bivalve larvae. *PLoS ONE*, 10(6), e0128376. <https://doi.org/10.1371/journal.pone.0128376> [WC]
- Walkusz, W., Williams, W. J., & Kwasniewski, S. (2013). Vertical distribution of mesozooplankton in the coastal Canadian Beaufort Sea in summer. *Journal of Marine Systems*, 127, 26–35. <https://doi.org/10.1016/j.jmarsys.2012.01.001> [ARC]
- Wallace, E. J., Looney, L. B., & Gong, D. (2018). Multi-decadal trends and variability in temperature and salinity in the Mid-Atlantic Bight, Georges Bank, and Gulf of Maine. *Journal of Marine Research*, 76(5), 163–215. [MAB]
- Wallace, R. B., Baumann, H., Grear, J. S., Aller, R. C., & Gobler, C. J. (2014). Coastal ocean acidification: The other eutrophication problem. *Estuarine, Coastal and Shelf Science*, 148, 1–13. <https://doi.org/10.1016/j.ecss.2014.05.027> [FLC]
- Wang, H. (2016). On shelf-slope water mass exchanges near Washington Canyon and Norfolk Canyon in the Mid-Atlantic Bight. Thesis, Master of Science, College of William & Mary, Virginia Institute of Marine Science, Paper 1539617966. <https://doi.org/10.25773/v5-jxbj-0a48> [MAB]
- Wang, Z. A., Wanninkhof, R., Cai, W.-J., Byrne, R. H., Hu, X., Peng, T.-H., & Huang, W.-J. (2013). The marine inorganic carbon system along the Gulf of Mexico and Atlantic coasts of the United States: Insights from a transregional coastal carbon study. *Limnology and Oceanography*, 58(1), 325–342. <https://doi.org/10.4319/lo.2013.58.1.0325> [SAG][MAB]
- Wang, Z. A., Lawson, G. L., Pilskaln, C. H., & Maas, A. E. (2017). Seasonal controls of aragonite saturation states in the Gulf of Maine. *Journal of Geophysical Research: Oceans*, 122(1), 372–389. <https://doi.org/10.1002/2016JC012373> [NE]
- Wanninkhof, R., Barbero, L., Byrne, R., Cai, W.-J., Huang, W.-J., Zhang, J.-Z., et al. (2015). Ocean acidification along the Gulf Coast and East Coast of the USA. *Continental Shelf Research*, 98, 54–71. <https://doi.org/10.1016/j.csr.2015.02.008> [SAG][MAB][NE]
- Wanninkhof, R., Pickers, P. A., Omar, A. M., Sutton, A., Murata, A., Olsen, A., et al. (2019). A surface ocean CO<sub>2</sub> reference network, SOCONET and associated marine boundary layer CO<sub>2</sub> measurements. *Frontiers in Marine Science*, 6, 400. <https://doi.org/10.3389/fmars.2019.00400> [WC]
- Wanninkhof, R., Pierrot, D., Sullivan, K. F., Barbero, L., & Triñanes, J. A. (2020). A 17-year dataset of surface water fugacity of CO<sub>2</sub>, along with calculated pH, Aragonite saturation state, and air-sea CO<sub>2</sub> fluxes in the Caribbean Sea. *Earth System Science Data Discussion*. <https://doi.org/10.5194/essd-2019-245> [NAT]
- Weijerman, M., Fulton, E. A., Kaplan, I. C., Gorton, R., Leemans, R., Mooij, W. M., & Brainard, R. E. (2015). An integrated coral reef ecosystem model to support resource management under a changing climate. *PLoS ONE*, 10(12), e0144165. <https://doi.org/10.1371/journal.pone.0144165> [PAC]
- Weinberg, J. R. (2005). Bathymetric shift in the distribution of Atlantic surfclams: Response to warmer ocean temperature. *ICES Journal of Marine Science*, 62(7), 1444–1453. [MAB]
- Weisberg, R. H., Liu, Y., Lembke, C., Hu, C., Hubbard, K., & Garrett, M. (2019). The coastal ocean circulation influence on the 2018 West Florida Shelf *K. brevis* red tide bloom. *Journal of Geophysical Research: Oceans*, 124(4), 2501–2512. <https://doi.org/10.1029/2018jc014887> [SAG]
- Whiteley, N. M. (2011). Physiological and ecological responses of crustaceans to OA. *Marine Ecology Progress Series*, 430, 257–271. [FLC]
- Wilkinson, M., Dumontier, M., Aalbersberg, I., Appleton, G., Axton, M., Baak, A., et al. (2016). The FAIR Guiding Principles for scientific data management and stewardship. *Scientific Data*, 3, 160018. <https://doi.org/10.1038/sdata.2016.18> [NAT]

- Williams, C. R., Dittman, A. H., McElhany, P., Busch, D. S., Maher, M. T., Bammler, T. K., & Gallagher, E. P. (2019). Elevated CO<sub>2</sub> impairs olfactory-mediated neural and behavioral responses and gene expression in ocean-phase Coho salmon (*Oncorhynchus kisutch*). *Global Change Biology*, 25(3), 963–977. <https://doi.org/10.1111/gcb.14532> [WC][INTRO]
- Williams, I. D., Baum, J. K., Heenan, A., Hanson, K. M., Nadon, M. O., & Brainard, R. E. (2015). Human, oceanographic and habitat drivers of central and western Pacific coral reef fish assemblages. *PLoS ONE*, 10(4), e0120516. <https://doi.org/10.1371/journal.pone.0120516> [PAC]
- Williams, N. L., Juranek, L. W., Feely, R. A., Johnson, K. S., Sarmiento, J. L., Talley, L. D., et al. (2017). Calculating surface ocean pCO<sub>2</sub> from biogeochemical Argo floats equipped with pH: An uncertainty analysis. *Global Biogeochemical Cycles*, 31(3), 591–604. <https://doi.org/10.1002/2016GB005541> [OO][WC]
- Wisshak, M., Schönberg, C. H. L., Form, A., & Freiwald, A. (2012). Ocean acidification accelerates reef bioerosion. *PLoS ONE*, 7(9), e45124. <https://doi.org/10.1371/journal.pone.0045124> [FLC]
- Wolf, D., Georgic, W., & Klaiber, H. A. (2017). Reeling in the damages: Harmful algal blooms' impact on Lake Erie's recreational fishing industry. *Journal of Environmental Management*, 199, 148–157. <https://doi.org/10.1016/j.jenvman.2017.05.031> [GL]
- Wongbusarakum, S., & Loper, C. (2011). Indicators to assess community-level social vulnerability to climate change: An addendum to SocMon and SEM-Pasifika regional socioeconomic monitoring guidelines (CRCP & TNC) <http://socmon.org/download.ashx?docid=64623> [FLC]
- Xu, Y.-Y., Cai, W.-J., Gao, Y., Wanninkhof, R., Salisbury, J., Chen, B., et al. (2017). Short-term variability of aragonite saturation state in the central Mid-Atlantic Bight. *Journal of Geophysical Research: Oceans*, 122(5), 4274–4290. <https://doi.org/10.1002/2017JC012901> [MAB]
- Xue, Z., He, R., Fennel, K., Cai, W.-J., Lohrenz, S., Huang, W.-J., Tian, H., et al. (2016). Modeling pCO<sub>2</sub> variability in the Gulf of Mexico. *Biogeosciences*, 13(15), 4359–4377. <https://doi.org/10.5194/bg-13-4359-2016> [SAG]
- Yamamoto-Kawai, M., McLaughlin, F. A., Carmack, E. C., Nishino, S., & Shimada, K. (2009). Aragonite undersaturation in the Arctic Ocean: Effects of ocean acidification and sea ice melt. *Science*, 326(5956), 1098–1100. <https://doi.org/10.1126/science.1174190> [ARC]
- Zhai, L., Platta, T., Tang, C. S., Sathyendranath, S., Fuentes-Yaco, C., Devred, E., & Wu, Y. (2010). Seasonal and geographic variations in phytoplankton losses from the mixed layer on the Northwest Atlantic Shelf. *Journal of Marine Systems*, 80, 36–46. [OO]
- Zhang, Y., Yamamoto-Kawai, M., & Williams, W. J. (2020). Two decades of ocean acidification in the surface waters of the Beaufort Gyre, Arctic Ocean: Effects of sea ice melt and retreat from 1997–2016. *Geophysical Research Letters*, 47, e60119. <https://doi.org/10.1029/2019GL086421> [OO]